

1 **Novel cell- and stage-specific transcriptional signatures defining *Drosophila* neurons, glia**
2 **and hemocytes**

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20 **Abstract:**

21 Cell types can be now defined at unprecedented resolution using high throughput assays. We
22 analyzed the transcriptional signatures of *Drosophila* neurons, glia and hemocytes, as examples of
23 cell types that are related by position (glia/neurons) or function (glia/hemocytes) or that are
24 unrelated (neurons/hemocytes). The most related cells display the highest similarity level (neurons
25 and glia), the least related ones, the lowest (neurons and hemocytes), however, cells can show
26 plastic features. Glia are much more similar to neurons than to hemocytes in the embryo, but are
27 equally similar to the two cell types in the larva, when hemocytes acquire more immune functions.
28 Larval glia and hemocytes display common as well as specific immune features, such as the glia-

29 specific NimA receptor, in agreement with the different environment faced by each cell types.
30 Surprisingly, time represents a key identity parameter, as neurons, hemocytes and glia group more
31 significantly by the stage than by the cell type and larval cells show upregulation of genes involved
32 in chromatin organization and in DNA repair. This latter group of genes is linked to changes in
33 gene expression levels and chromatin organization, revealing a function of these genes beyond
34 DNA repair. Finally, the metabolic profiles reveal cell type-specific signatures and an overall shift
35 from an embryonic, anabolic state to a larval, catabolic state.

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37

38 **Introduction**

39 Traditionally, cell types have been defined by morphology, position, origin and function, relying
40 on the expression of few proteins. While the combination of these parameters has produced
41 valuable information, each of them on their own can lead to misleading conclusions. Cells in
42 different organs can display similar functions and cells within the same organ can have different
43 properties. Moreover, the *a priori* selection of proteins can introduce bias and neglect the wealth
44 of molecular information intrinsic to each cell type. Finally, functional plasticity and dynamic
45 states may conduct to a reductionist classification. To ensure a more unbiased and quantitative
46 analysis, we used a high throughput approach and a data-driven computational method on neurons,
47 hemocytes and glia. To account for plasticity, we analyzed them at two stages, embryo and larva.

48 *Drosophila* neurons and glia differentiate from common, ectodermal, precursors. Neurons sense,
49 process and transmit information or secrete neuropeptides (Garces and Thor, 2006; Landgraf and
50 Thor, 2006; Schmid et al., 1999). Glia are involved in nervous system development, function, and
51 maintenance. They control cell proliferation (Ebens et al., 1993), axonal and synapse development
52 (Brink et al., 2012; Ou et al., 2014), neuronal insulation and survival (Shepherd, 2000; Stork et al.,
53 2008; Volkenhoff et al., 2015). They also establish the blood-brain barrier (BBB) (Abbott, 2005)
54 and provide the immune function within the nervous system (Awasaki and Ito, 2004; Sonnenfeld
55 and Jacobs, 1995a; Watts et al., 2004). The mesodermally derived hemocytes patrol the organism
56 to ensure cellular immunity outside the nervous system (Lebestky et al., 2000; Lemaitre et al.,

57 1996; Tepass et al., 1994), hence sharing the function but not the origin with glial cells, while
58 sharing neither the origin nor the function with neurons.

59 Transcriptome comparison highlights cell-specific features. Neurons and hemocytes show the least
60 related transcriptional landscapes regardless of the stage. Glia and neurons stably share the highest
61 similarity, in line with the fact that they constantly interact to ensure the homeostasis of the nervous
62 system. Glia and hemocytes display significant similarity in the larva. This goes along with a
63 significant change of the hemocyte transcriptional profile from the embryo to the larva, an open
64 system that has to react to external stimuli as pathogens. Although larval glia and hemocytes share
65 an immune function, the two cell types encounter distinct environments, pathogens for hemocytes,
66 axon and dendrite debris for glia. Accordingly, the Eater and the Nimrod C1 (NimC1) scavenger
67 receptors are specific to hemocytes, NimA is specific to glia.

68 Similarly, some metabolic features are common, others are specific with respect to tissue and stage.
69 The key glycolytic gene *lactate dehydrogenase (ldh)* is upregulated in all the three cell types in
70 the embryo, down-regulated in the larva. The expression of lipogenic genes, on the other hand, is
71 upregulated in all three embryonic cell types, but changes discretely as the embryo transitions to
72 the larval stage. Hemocytes dramatically shift to a lipolytic state while neurons maintain their
73 lipogenic state. Larval glia maintain the expression of lipogenic genes but shows additional
74 upregulation of lipolytic pathway genes. The observed common metabolic states may be a
75 reflection of the overall developmental program, but the cell-specific states may highlight
76 metabolic needs necessary to accommodate distinct functional roles.

77 Transcriptome comparison also highlights common stage-specific features. Larval cells express
78 genes controlling chromatin organization as well as DNA repair at high levels. One of the main
79 DNA repair genes is *rad50*, involved in DNA double strand break (DSB) repair. Its depletion
80 triggers a genome-wide change in chromatin organization and mis-regulation of gene expression,
81 pointing to a role beyond DSB. Finally, the three cell types display a glycolytic and lipogenic state
82 in the embryo, a lipolytic and oxidative state in the larva, changes that are likely linked to external
83 inputs such as feeding and respiration.

84 Altogether, our genome-wide analysis makes it possible to disentangle the relative importance of
85 origin, position, function and time in the definition of cell types. The expression profile of a cell
86 population is strongly affected by their origin in the embryo, while larval cells acquire additional

87 levels of specialization and become more affected by the function. Moreover, the three cell types
88 display different degrees of plasticity, the migratory hemocytes exposed to the external challenges
89 being the most plastic ones. This work paves the way for systematic identification of gene networks
90 and expression programs, which will ultimately shed light on the biological mechanisms
91 characterizing cell diversity in multicellular organisms.

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93

94 **Results**

95 **Impact of origin, stage and function on the transcriptional landscapes**

96 To assess the impact of origin, function and position on the definition of a cell type, we compared
97 the transcriptomes of neurons and glia, which share the origin; those of hemocytes and glia, which
98 share the scavenging activity; and those of neurons and hemocytes, which share neither origin nor
99 function. We performed the comparison in mature embryos (stage 16 or E16) and in wandering
100 third instar larvae (L3), to understand whether expression profiles change from a small, closed and
101 immobile system to a bigger, open system that is able to grow and to respond and adapt to the
102 environment. Glia, hemocytes and neurons were purified from transgenic lines specific to each
103 cell type: *repo-nRFP*, *srp(hemo)Gal4/+*; *UAS-RFP/+* and *elav-nRFP*, respectively. The
104 hierarchical clustering of the transcriptomes shows a good correlation between the biological
105 replicates, highlighting a low biological variability (**Figure 1-a**).

106 The developmental stage is the main factor of heterogeneity, more important than the cell type, as
107 the highest correlation in the dendrogram is seen for samples of the same stage. Thus, each cell
108 type is more similar to the two others at the same stage than to the same cell type at a different
109 stage. The dendrogram also shows that glia and neurons group together, sharing the highest degree
110 of similarity. We next performed pairwise comparisons of the number of differentially expressed
111 genes to evaluate the commonalities and the differences between neurons, glia and hemocytes. We
112 defined significantly upregulated genes based on transcripts that are enriched in one population
113 compared to the other following three parameters: number of reads > 15, adjusted p-value <0.05
114 and fold change >1.5 between the number of reads. Upregulated genes in each cell type were
115 compared in the Venn diagram. The comparison of the cell-specific transcriptomes is shown in

116 (Figure 1-b) for the embryo, in (Figure 1-c) for the larva. The highest number of shared
117 upregulated genes concerns E16 neurons and glia (in green) (31.3% of all genes included in the
118 diagram), while only 5.7% of shared upregulated genes are found between E16 hemocytes and glia
119 (in orange). Interestingly, the shared enriched transcripts between hemocytes and glia reach 18.3%
120 by the larval stage, suggesting that glia and hemocytes acquire common molecular features by L3,
121 despite their distinct origin. Finally, the Venn diagram and the dendrogram indicate that glia and
122 neurons maintain a high degree of similarity in the larva (27.1%). Neurons and hemocytes remain
123 the most unrelated cell populations, with 5.9% of shared enriched transcripts at E16, 3,8% at L3.

124 Interestingly, the number of expressed genes is overall lower in L3 compared to E16, and this is
125 true for all cell types (transcripts with at least 15 reads, Figure 1-d,e, top three lines). By contrast,
126 the number of upregulated genes in each cell type increases by L3 (Figure 1-d,e, bottom three
127 lines). 10838 genes are expressed in E16 glia, 11281 in E16 neurons and 9811 in E16 hemocytes.
128 By L3, 9187 genes are expressed in glia, 9312 in neurons and 7828 in hemocytes, which means
129 that at least 1500 genes are only expressed at E16 in each cell type compared to L3 (Figure 1-d,e).
130 This is not due to a low sequencing depth, since the same number of cells was used for each sample
131 and there are no differences between the number of reads mapped for embryonic and larval cells
132 (Sup. Figure 1-a). To further understand the molecular mechanisms underlying the stage
133 differences, we analyzed the expression profiles of transcription factors, proteins that coordinately
134 control downstream effector genes. In line with the increased number of cell-specific upregulated
135 genes in the larva, the number of transcription factors commonly expressed in all three cell types
136 decreases by L3 (Sup. Table 1-a,b). Thus, differentiated cells become more specialized over time.

137 In sum, our results show that the developmental stage has a stronger impact on the transcriptome
138 than the cell type. In agreement with their respective origin and position, glia and neurons display
139 the strongest similarity, hemocytes and neurons display the opposite behavior. Glia and hemocytes
140 become more similar by L3. Finally, larval differentiated cells express fewer genes than the
141 embryonic counterparts, but, proportionally, more genes that are upregulated in a cell-specific
142 manner.

143

144 **Cell-specific signatures in the embryo define neurons and glia but not hemocytes**

145 The comparison of the transcriptomes indicates that almost half of the transcripts are common to
146 neurons, glia and hemocytes (see **Figure 1-d,e**: number of genes expressed – number of genes
147 upregulated in each cell type), calling for more refined parameters to define the three cell types.
148 Since a significant fraction is enriched only in one cell type or shared by two cell populations, we
149 predicted that a pairwise comparison associated with quantitative and Gene Ontology (GO)
150 analyses would allow the disclosure of the signatures and functions that better define cell types.

151 In the embryo, 1649 (out of 11281) genes are upregulated in neurons compared to glia (in blue),
152 992 in glia compared to neurons (out of 10838) (in yellow) (**Figure 2-a**). Thus, relatively few
153 genes are upregulated in a specific cell type. The vast majority of the genes are non-differentially
154 expressed, with number of reads > 15 and adjusted p-value of fold change > 0.05. On average,
155 these genes are expressed at lower levels than those upregulated in a specific cell type (**Sup. Figure**
156 **1-b,c**). Amongst the non-differentially expressed genes, the ones with fold change < 1.5 were
157 considered as commonly expressed, mostly involved in biological processes found in all cells such
158 as ncRNA processing, ribosome biogenesis and mRNA metabolic process (**Sup. Figure 1-d, Sup.**
159 **Table 2-a**). The transcripts enriched in neurons are mostly involved in neuronal pathways, such
160 as synaptic transmission, neurotransmitter secretion and neuron projection guidance (**Figure 2-b,**
161 **Sup. Table 2-b**). Those enriched in glia are involved in the establishment of the BBB, axon
162 ensheathment and the development of glial cells (**Figure 2-c, Sup. Table 2-c**).

163 The comparative analysis between E16 glia and hemocytes reveals a much higher number of
164 transcripts specifically enriched in either cell type. This is in line with the Venn diagram in **Figure**
165 **1-b**. 1945 transcripts are enriched in hemocytes (in red) and 2937 in glia (in yellow) (**Figure 2-d**).
166 Interestingly, the hemocyte enriched transcripts only highlight one GO term related to immunity,
167 ‘positive regulation of immune functions’, which comprises 39 genes (**Figure 2-e, Sup. Table 2-**
168 **d**). Among them are 14 members of the Toll pathway and 6 of the Immune deficiency (Imd)
169 pathway (**Figure 2-g**). The Imd pathway is one of the conserved NF- κ B (nuclear factor kappa-
170 light-chain-enhancer of activated B cells) immune signaling pathways in insects that regulates an
171 antibacterial defense response. The remaining GO terms include “macroautophagy” and “Golgi
172 vesicle transport”. In the case of glia, we find glia-specific functions such as septate junction

173 assembly (involved in BBB establishment), glial cell differentiation and neural functions such as
174 motoneuron axon guidance (Abbott, 2005; Barres, 2008) (**Figure 2-f, Sup. Table 2-e**).

175 The comparison between E16 neurons and hemocytes also shows poor similarity, with 5261 genes
176 non differentially expressed, 3608 upregulated in neurons, 1990 in hemocytes (**Figure 2-h**).

177 Altogether, these data reveal that only a minority of transcripts is cell-specific and that embryonic
178 glia and neurons are closer to each other than to hemocytes. The genes that are specifically
179 upregulated in E16 recapitulate the specific functions of neurons and glia, while few cell-specific
180 terms are identified in hemocytes.

181

182 **Over development, hemocytes display higher plasticity compared to neurons and glia**

183 Cell functions adapt to changing environments during the life cycle. Typically, embryos live in
184 autarchy, whereas larvae function as open systems, due to feeding, respiration and exposure to
185 pathogens. Accordingly, although already functional in the embryo, we found that the
186 differentiated cells become increasingly specialized by the larval stage.

187 A pairwise comparison reveals that 2500 genes are upregulated in L3 neurons compared to glia
188 and 2279 genes are upregulated in L3 glia compared to neurons (**compare Figure 3-a and Figure**
189 **2-a**). The genes upregulated in L3 neurons highlight neuronal functions and molecular cascades
190 linked to signaling (neuropeptide signaling pathway, regulation of neurotransmitter levels and
191 synapse organization, **Figure 3-b, Sup. Table 3-a**), compared to the GO terms linked to
192 development found at E16 (**Figure 2-b**). L3 glia display the GO term septate junction assembly
193 also found at E16, likely due to the fact that the BBB starts forming in the embryo and continues
194 to expand as the CNS grows during the larval stages (reviewed in (Limmer et al., 2014). In
195 addition, terms related to the differentiation of glia and gliogenesis were also found (**Figure 3-c,**
196 **Sup. Table 3-b**), in line with the extensive glial proliferation ongoing in the larva (Pereanu et al.,
197 2005).

198 We performed the pairwise comparison between L3 hemocytes and glia. Unlike their embryonic
199 counterpart, L3 hemocytes present several terms related to immune functions such as
200 phagocytosis, positive regulation of antimicrobial humoral response and positive regulation of

201 immune effector process (**compare Figure 2-e and Figure 3-e, Sup. Table 3-c**). The clear shift
202 in the hemocyte expression profile indicates a major change in the function of these cells during
203 development. The genes upregulated in glia (2941 genes in yellow) are involved in glial functions
204 such as synapse organization and axon guidance (**Figure 3-f, Sup. Table 3-d**), a function that has
205 already been investigated (reviewed in (Bittern et al., 2021)).

206 The MA plot comparing larval neurons and hemocytes shows that these cell types display the
207 lowest level of similarity, comparable to that observed in the embryo (**Figure 3-g**).

208 The comparative analyses show that the divergence between neurons and glia increases over time:
209 higher number of upregulated genes at L3 compared to E16 (Compare **Figure 3-a to Figure 2-a**),
210 increased fold change of upregulated genes at L3 (**Sup. Figure 2-a**). By contrast, in the case of
211 hemocytes and glia, the number of upregulated genes as well as their median fold change value
212 are similar in E16 and L3 (compare **Figure 2-d and Figure 3-d, Sup. Figure 2-a**).

213 To shed light on the hemocyte / glia similarities at L3, we identified the genes that are upregulated
214 in L3 glia and hemocytes but not in neurons (**Figure 1-c genes in orange**). A GO term analysis
215 highlights genes involved in immune processes and in metabolism (**Sup. Table 4-a**). Typically, 4
216 genes are involved in fatty acid oxidation including CPT2, Mtpalpha, Echs1, Mfe2, CG4860 and
217 CG17544 (**Sup. Figure 2-b, Sup. Table 4-b**), indicating the significant reliance of L3 glia and
218 hemocytes on lipid break-down. Moreover, the upregulated genes in the L3 hemocytes and glia
219 show several GO terms related to immune functions such as positive regulation of antimicrobial
220 humoral response and positive regulation of immune effector process (**Sup. Figure 2-b, Sup.**
221 **Table 4-a,c**).

222 To better understand the nature of this evolving cell behavior, we compared each cell type at E16
223 and L3. The MA plots show that neurons and glia evolve in a similar manner: a comparable number
224 of genes is non-differentially expressed in neurons and in glia between E16 and L3; also, many
225 more genes are upregulated in E16 glia and neurons, than in L3 glia and neurons (**Sup. Figure 2-**
226 **c,d**). By comparison, the number of non-differentially expressed genes in E16 and L3 hemocytes
227 is significantly lower than that detected in glia and neurons (**Sup. Figure 2-e**). This highlights the
228 robust shift in the hemocytes' transcriptome during development compared to the relatively less
229 dynamic transcriptomes of neurons and glia.

230 Finally, and in agreement with the gene expression profiles, neurons and glia share the highest
231 number of transcription factors at E16 (**Sup. Figure 2-f, Sup. Table 1-c,d,e**), hemocytes and glia
232 at L3 (**Sup. Figure 2-g, Sup. Table 1-f,g,h**), further highlighting the increased commonality
233 between hemocytes and glia at L3.

234 In sum, cell-specific features and specialization increase over time, especially for hemocytes,
235 which acquire a stronger immune potential and share more similarities with glia in the larva.

236

237 **Glia and hemocytes express specific scavenger receptors**

238 The vast majority of the hemocytes (95%) act as scavenger cells in homeostatic and challenged
239 conditions. In the embryo, they are necessary for the elimination of apoptotic bodies and for the
240 clearance of cellular debris. Prior to the formation of the BBB, they also contribute to this function
241 within the developing CNS (Franc et al., 1996; Kurant et al., 2008; Manaka et al., 2004; Sonnenfeld
242 and Jacobs, 1995b). Once glial cells form the BBB, however, hemocytes no longer have access to
243 the CNS and by late embryogenesis the scavenging function within the nervous system is taken up
244 by glia solely (Kurant et al., 2008). Thus, hemocytes and glia represent respectively the
245 macrophages acting outside and inside the nervous system. The question then arises as to whether
246 glia express the same immune pathways of hemocytes or represent a distinct class of macrophages.

247 A key molecular signature of macrophages is the expression of scavenger receptors (Kocks et al.,
248 2005; Kurucz et al., 2007; Lebestky et al., 2000; Lemaitre et al., 1996; R met et al., 2002; Tepass
249 et al., 1994), which we therefore used to compare hemocytes and glia. The MA plots comparing
250 E16 (**Figure 4-a, Sup. Table 5-a**) or L3 hemocytes and glia (**Figure 4-b, Sup. Table 5-b**) allowed
251 us to subdivide the receptors based on their relative levels of expression and fold change between
252 the two cell types. Group 1 is specific to hemocytes, group 2 is expressed in both cell types, group
253 3 is specific to glia (**Figure 4-a,b group 1 in red, group 2 in black, group 3 in yellow**).

254 Group 1 receptors are involved in the phagocytosis of Gram-positive and Gram-negative bacteria
255 (Pearson et al., 1995; R met et al., 2001), matching the hemocyte-specific function of pathogen
256 phagocytosis. They include the well-studied genes *eater*, *NimC1*, *NimC2* and *Scavenger receptor*
257 *class C, type I (Sr-CI)*. *eater*, *NimC1* and *NimC2* contain multiple epidermal growth factor (EGF)
258 like repeats called NIM repeats (Kurucz et al., 2007). SR-CI and SR-CIV belong to the *Drosophila*

259 SR-C class of scavenger receptors, SR-CI contains several known motifs, including two
260 complement control protein domains, a somatomedin B domain and a Meprin, A-5 protein, and
261 receptor protein-tyrosine phosphatase Mu domain (Rämet et al., 2001).

262 Group 2 receptors include *NimC4*, *draper (drpr)* and *croquemort (crq)*, which was believed to be
263 only expressed in hemocytes (Franc et al., 1996) and belongs to the SR-B class of scavenger
264 receptors homologous to the mammalian CD36 (Franc et al., 1999). Although *crq* is expressed at
265 higher levels in hemocytes, it is still expressed in glia at relatively high levels and cannot be
266 considered hemocyte-specific (**Sup. Table 5-a,b**). These receptors are involved in the
267 phagocytosis of apoptotic bodies and debris, in line with the fact that *Drpr* and *NimC4* act in
268 hemocytes and in glia to promote the clearance of apoptotic cells and axonal debris (Kurant et al.,
269 2008; MacDonald et al., 2006; Manaka et al., 2004; Melcarne et al., 2019; Roddie et al., 2019).

270 Group 3 includes three genes: *bark beetle (bark)* encodes a transmembrane scavenger receptor-
271 like protein important for the establishment of the BBB (Hildebrandt et al., 2015; Parker and Auld,
272 2006; Stork et al., 2008). *Secreted Wg-interacting molecule (Swim)* contains a somatomedin B
273 domain similar to Sr-CI, however it has not been investigated for its role in phagocytosis. *NimA*
274 is a *Drpr*-like receptor containing one NIM domain followed by two EGF repeats and one Emilin
275 domain (**Sup. Figure 3-a**, Callebaut et al., 2003). Its *C. elegans* orthologs are involved in the
276 phagocytosis of apoptotic cells (Kurucz et al., 2007; Mangahas and Zhou, 2005).

277 The expression profile of the receptors evolves with time. The number of scavenger receptors
278 specific to hemocytes and their expression levels increases in the larva (4 at E16, 8 at L3, shown
279 in red, **Figure 4-a,b**). In parallel, the number of receptors non-differentially expressed between
280 hemocytes and glia decreases in the larva (8 at E16, 5 at L3, shown in black). Moreover, within
281 the same cell type, the expression levels of some receptors change significantly, *eater* being highly
282 upregulated in L3, *NimC4* at E16.

283 To validate the expression of these genes, we performed quantitative reverse transcription PCR
284 (RT-qPCR) assays on L3 glia and hemocytes. We tested and confirmed the purity of the sorted
285 cell populations by quantifying the expression levels of *Hemolectin (Hml)* and *reverse polarity*
286 (*repo*), as markers of hemocytes and glia, respectively (**Sup. Figure 3-b,c**). *NimC1* and *eater* are
287 significantly upregulated in L3 hemocytes compared to glia (**Sup. Figure 3-d,e**), *drpr* shows

288 comparable expression levels (**Sup. Figure 3-f**); *crq* seems to be more expressed in hemocytes
289 however the difference is not statistically significant (**Sup. Figure 3-g**), similarly to the
290 transcriptome results. *NimA* is only found in glia (**Sup. Figure 3-h**). The finding that some
291 receptors are expressed in glia and hemocytes, whereas others are cell-specific recapitulates their
292 common function of phagocytosing apoptotic cells and debris and at the same time emphasizes the
293 role of hemocytes in host defense against pathogens. In addition, the higher levels of the scavenger
294 receptors in L3 than in E16 hemocytes prove an increased functional specificity of hemocytes in
295 the larva.

296 Given the high specificity and levels of expression of *NimA*, we performed an *in situ* hybridization
297 RNAscope assay on wild type (*WT*) L3 CNS and found very specific labeling at the position of
298 the neuropile (**Sup. Figure 3-i,i'**). We complemented this analysis by labeling *NimA-Gal4; UAS-*
299 *eGFP* E16, L3 and adults with Repo, a pan-glial marker and Nazgul, a NADP-retinol
300 dehydrogenase specifically expressed in embryonic astrocyte and ensheathing glia, the two cell
301 types that constitute the neuropile glia (Peco et al., 2016; Ryglewski et al., 2017) (**Figure 4-c-e'**).
302 Nazgul and *NimA* nicely colocalize in the embryonic glial cells (**Figure 4-c,c'**). By contrast, *NimA*
303 and Nazgul are found in larval glial cells that surround the neuropile and are in close proximity,
304 suggesting that at that stage Nazgul is expressed in the astrocyte glia that invade the neuropile,
305 while *NimA* is expressed in the ensheathing glia as well as in the wrapping glia that surround the
306 peripheral nerves (**Figure 4-d,d'**). In the adult CNS, *NimA* is also found in glial cells surrounding
307 the neuropile and, based on single-cell RNA seq data, *NimA* is expressed in adult ensheathing glia
308 but not in astrocyte-like glia (**Figure 4-f,f'**) (Davie et al., 2018). Thus, *NimA* represents a novel
309 glia-specific scavenger receptor expressed in a subset of glia cells throughout the life of the fly.

310 In sum, hemocytes have a strong commitment to immune functions in L3 and each phagocytic cell
311 type expresses common as well as specific receptors, such as the glia-specific *NimA* gene and the
312 hemocyte-specific *NimC1* and *eater* genes. This highlights a cell-specific phagocytic potential that
313 differentiates glia from the macrophages outside the CNS.

314

315 **Stage-specific transcriptional signatures**

316 The transition from the embryo to the larva involves massive physiological changes that affect the
317 organism as a whole. This likely explains why the transcriptomes group by stage more than cell
318 types (**Figure 1-a**). To get more mechanistic insights on the changes associated with time, we
319 assessed whether a stage-specific molecular signature is common to all cell types. We combined
320 all cells deriving from E16 and compared their transcriptomes to those of cells deriving from L3.

321 The MA plot comparing E16 to L3 transcriptomes suggests that embryonic cells are overall closer
322 to each other than larval cells (**Figure 5-a**), in line with the dendrogram in **Figure 1-a**. This does
323 hint to a stage-specific signature common to all cells. The transcripts enriched at E16 compared to
324 L3 are involved in cytoplasmic translation and cuticle development (**Figure 5-b, Sup. Table 6-a**).
325 Genes included in the GO term cytoplasmic translation code for ribosomal proteins and translation
326 machinery, such as translation initiation factors. Ribosomal protein transcripts and ribosomes are
327 known to be maternally deposited and highly abundant in early embryos (Qin et al., 2007), but
328 new ribosomes are built by the end of embryogenesis, in line with the transcriptomic data. The
329 ‘cuticle development genes’, include 42 genes coding for Cuticular proteins, 21 genes coding for
330 members of the Tweedle family and 7 genes coding for CPR cuticle protein family (**Figure 5-c,**
331 **Sup. Table 6-b**). These proteins are predicted to be present in the extracellular matrix (ECM)
332 (Cornman, 2009; Karouzou et al., 2007; Naba et al., 2016; Zuber et al., 2020), which implies a
333 role of all three cell types in the deposition of ECM and cuticle at embryonic stage, a feature that
334 has already been described for the hemocytes (Brown, 2011; Martinek et al., 2008) and is in
335 accordance with the important role of the ECM in organogenesis.

336 Fewer genes are upregulated in L3 compared to E16 (**Figure 5-a**) and many of them belong GO
337 terms related to tRNA aminoacylation, DNA repair and epigenetics (**Figure 5-d, Sup. Table 6-c**).
338 The enrichment of tRNA aminoacylation related transcripts hints to increased translation, which
339 could be due to the fact that, unlike embryos, larvae take up amino acids and nutrients through
340 feeding and are thus able to aminoacylate tRNAs, as lack of amino acids and starvation has been
341 shown to cause significant decreases in protein synthesis (Deliu et al., 2017).

342 We were puzzled by the enrichment of transcripts involved in DNA repair. Among them are *rad50*,
343 *Nijmegen breakage syndrome (nbs)* and meiotic recombination 11 (*mre11*), members of the *MRN*

344 complex, as well as *Telomere fusion (Tefu)*, the ortholog of *Ataxia Telangiectasia Mutated (ATM)*
345 **(Figure 5-e, Sup. Table 6-d)**. These genes have been implicated in DSB repair (Wang et al.,
346 2014), in DNA replication restart after replication stress (Gatei et al., 2014) and in preventing
347 telomere fusion as well as chromosome breakage in *Drosophila* (Ciapponi et al., 2004, 2006). To
348 validate the stage-specific enrichment of these transcripts, we performed qPCR assays on RNA
349 extracted from whole E16 and L3 animals: *rad50* **(Sup. Figure 4-a)** and *mre11* **(Sup. Figure 4-**
350 **b)** are upregulated in L3 and *nbs* tends to be upregulated **(Sup. Figure 4-c)**. Based on the fact that
351 double strand breaks trigger the phosphorylation of the histone H2AX, a variant of the H2A protein
352 family (Redon et al., 2002), we hypothesized that the increase in DNA repair genes is due to an
353 increase in DSBs. Therefore, we estimated the relative levels of DSB using the phosphorylated
354 H2A.v (H2A.v-P) *Drosophila* marker, the equivalent of gamma-H2AX in mammals. When DNA
355 damage forms double strand breaks, H2A.v is phosphorylated by kinases such as ATM and the
356 phosphorylated protein, gamma-H2AX, is the first step in recruiting and localizing DNA repair
357 proteins. Western blot assays on histone extracts show that the levels of H2A.v-P decrease in L3
358 compared to E16 **(Sup. Figure 4-d,e)**, contrary to our initial hypothesis.

359 A recent study showed that the yeast equivalent of the MRN complex (MRX) is involved in
360 controlling gene expression levels through modulating chromatin interaction (Forey et al., 2021).
361 Moreover, MRN components interact with Heterochromatin Protein 1 (HP1), which is involved in
362 gene repression through heterochromatin formation (James and Elgin, 1986; James et al., 1989),
363 and they regulate HP1 expression and stability (Bosso et al., 2019). This, in addition to the finding
364 of GO terms related to epigenetics “negative regulation of gene expression, epigenetic”,
365 “chromatin remodeling” and “histone modification” in the genes upregulated in L3 compared to
366 E16 **(Figure 5-f, Sup. Table 6-e)** led us to ask whether Rad50 might be involved in chromatin
367 conformation and in the regulation of gene expression.

368 Chromatin states are defined by a highly complex epigenetic code comprised of histone post-
369 translational modifications. Histone methylation is complex in that residues may be mono-, di-, or
370 trimethylated, and these marks may activate or repress gene transcription. Di- and trimethylation
371 of H3K4, for instance, strongly correlate with active gene transcription, whereas H3K9 and H3K27
372 methylation are mostly associated to a closed heterochromatin state and therefore to transcription
373 repression (Aranda et al., 2015; Conway et al., 2015; Peters et al., 2003; Rea et al., 2000; Santos-

374 Rosa et al., 2002). To determine whether Rad50 is required for chromatin conformation and
375 repression of gene expression in L3 larvae, we labeled polytene chromosomes of wild-type or
376 *rad50Δ5.1* mutant larvae with antibodies against H3K4me3, H3K9me2 or H3K9me3. The absence
377 of Rad50 causes a significant increase of H3K4me3 and a decrease of H3K9me2/me3 epigenetic
378 marks (**Figure 6-a,b**). Polytene chromosomes are characterized by the alternation of dense
379 (heterochromatic) and less dense (euchromatic) bands, a reflection of the chromatin condensation
380 state. Interestingly, we observed an alteration of the H3K4me3 band signal profile accompanied
381 by almost complete absence of H3K9me3 in *rad50Δ5.1* polytene chromosomes compared to *WT*
382 (**Figure 6-c,d**). While the repression marks H3K9me2/me3 show a clear decrease in *rad50Δ5.1*,
383 the H3K27me3 signal increases significantly in *rad50Δ5.1* (**Figure 6-e,f**). Due to the link between
384 H3K27me3 and DNA damage shown by studies highlighting an increase in H3K27me3 at DNA
385 DSB sites (Basenko et al., 2015; O'Hagan et al., 2008, 2011) and the role of Rad50 in DNA
386 damage, we asked whether the increase in H3K27me3 is simply due to increased DSBs.
387 Comparing H3K27me3 and H2A.v-P distribution shows a clear lack of colocalization of the two
388 marks thus highlighting that the increase in H3K27me3 is independent from the increase of DSBs
389 (**Figure 6-e-g**).

390 The changes in the H3K4me3 and H3K27me3 profiles and the strong reduction of H3K9me3,
391 observed in *rad50Δ5.1* polytene chromosomes, suggest that Rad50 depletion affects chromatin
392 organization and hence gene expression regulation. These findings, together with the published
393 data, underline a novel role of the DNA damage factors, in particular Rad50, in regulating
394 chromatin states.

395 To understand whether Rad50 depletion and its effect on chromatin state also impinge on gene
396 expression, we analyzed several transcripts that show high expression in E16 but low expression
397 in L3 in all cell types in the *WT*. Assessing the expression of these genes in L3 *WT* compared to
398 *rad50Δ5* showed significant upregulation of four genes upon Rad50 depletion including the
399 *TweedleW* (*TwdlW*), *ftz transcription factor 1* (*ftz-f1*), *Uncoordinated 76* (*Unc-76*) and the member
400 of ATM *Mre11* (**Figure 6-h,k**), while *olf413* and *snail* (*sna*) show significant decrease of
401 expression levels (**Figure 6-l,m**). Altogether, these and the published data highlight a novel role
402 of the DNA damage factors, and specifically Rad50, in transcriptional regulation.

403 In sum, our data reveal that different developmental stages are associated with specific molecular
404 signatures, regardless of the cell type, including chromatin organization.

405

406 **Metabolic signatures of embryonic and larval neurons, glia and hemocytes**

407 A careful comparison of the embryonic and larval cells also revealed distinct metabolic states.
408 Compared to their larval counterpart, E16 hemocytes express more glycolytic enzymes such as
409 *lactate dehydrogenase (ldh)*, essential for the conversion of pyruvate to lactate (Cattenoz et.al.,
410 2020, **Figure 7**). This implies the activation of a Warburg-like glycolytic state and the conversion
411 of pyruvate to lactate. E16 hemocytes are also lipogenic as they highly express *de novo* the
412 lipogenic enzyme, *Acetyl-CoA carboxylase (ACC)* and enzymes of the triacylglycerol biosynthetic
413 pathway (TAG) that include *Fatty acid synthase (FAS)*, *Glycerol-3-phosphate acyltransferase*
414 (*Gpat*), *1-Acylglycerol-3-phosphate O-acyltransferase (Agpat)*, *Lipin (Lpin)* and *Diacylglycerol*
415 *O-acyltransferase (DGAT)*. By contrast, L3 hemocytes are more oxidative, they most likely
416 internalize and metabolize lipids through the beta-oxidation pathway to generate acetyl CoA and
417 drive the tricarboxylic acid (TCA) cycle. This is supported by the enrichment of transcripts
418 encoding lipid-scavenging receptors and by the poor expression of genes involved in the TAG
419 pathway (**Figure 7, Sup. Table 7**). Moreover, L3 hemocytes do not rely on glycolysis to drive the
420 TCA cycle, as the genes involved in glycolysis, mainly *phosphofructokinase* and *pyruvate*
421 *dehydrogenase (PDH)*, of which *PDH* controls the rate-limiting conversion of pyruvate to acetyl
422 CoA and critically controls TCA activity, are not upregulated in those cells. Finally, the poor
423 expression of *phosphoenolpyruvate carboxykinase (PEPCK)* and *fructose 1,6-bisphosphatase*
424 (*FBPase*), as well as a significant enrichment of the *Glut1* sugar transporter transcripts suggest the
425 absence of gluconeogenesis and the active uptake of glucose in the larva. Because the glycolytic
426 genes downstream of *glucose 6, phosphate (G6P)* are poorly expressed while those of the pentose
427 phosphate pathway (PPP) are highly expressed in the L3 hemocytes, the internalized glucose could
428 be used to generate pentose sugars for ribonucleotide synthesis and redox homeostasis through the
429 generation of NADPH. Hence, apart from the glycolytic pathway, E16 and L3 hemocytes display
430 anabolic (gluconeogenic, lipogenic) and catabolic (oxidative, lipolytic) signatures, respectively.

431 Comparative analyses revealed interesting commonalities in a few metabolic pathways, in addition
432 to metabolic changes that are cell-specific, which projects underlying differences in the metabolic
433 states (**Figure 7**). The one feature observed across all cell types is a clear metabolic shift in the
434 key glycolytic enzyme *ldh*, which is highly expressed in the embryo (**Figure 7, Sup. Table 7**).
435 This implies a global activation of a Warburg like state specifically in the embryo.

436 The transcripts of the glycolytic pathway genes are enriched in E16 glia and neurons: *Glucokinase*
437 (*Hexokinase-C*, *FBgn0001187*), *Aldolase* and *Glyceraldehyde 3-phosphate dehydrogenase*
438 (*Gapdh*) show prominent expression in glia, while only *Glucokinase* and *pyruvate kinase* are
439 expressed at high levels in the neurons. The transcripts coding for the *Glucose 6-phosphate*
440 *dehydrogenase (G6PD)*, a rate-limiting enzyme of the PPP pathway, is also strongly enriched in
441 E16 glia and neurons. This implies that these cells most likely use glucose for driving the PPP
442 pathway and glycolysis. Neither E16 glia nor E16 neurons are gluconeogenic, as the *PEPCK* gene
443 is poorly expressed at that stage. We therefore assessed the underlying source of glucose driving
444 glycolysis and evaluated the capacity for glucose uptake via transporters as a predictable source.
445 This revealed a strong enrichment of transcripts coding for sugar transporters like *sut1*, *sut3*, *sut4*
446 and specifically glucose transporters *Glut1* and *Glut3*. L3 glia and neurons seem both
447 gluconeogenic and glycolytic. Thus, E16 glia and neurons rely on extracellular sugar uptake,
448 sustained glycolysis and PPP while L3 glia and neurons utilize *de novo* gluconeogenesis to fuel
449 glycolysis.

450 The fate of glucose, post its breakdown to pyruvate via glycolysis, apart from driving lactate, can
451 be diverted to the TCA cycle (oxidative). We did observe a significant upregulation the TCA
452 enzymes in E16 glia compared to their larval counterpart. For instance, *Citrate synthase*,
453 *Aconitase*, *Isocitrate dehydrogenase (IDH)*, *oxoglutarate dehydrogenase (Ogdh)*, *succinate*
454 *dehydrogenase (SDH)* and *malate dehydrogenase (MDH)* are highly expressed in the embryo. By
455 contrast, only a subset of TCA enzymes (*Aconitase*, *succinyl CoA synthetase (SCS)*, *Fumarate*
456 *hydratase (FH)*) show prominent expression in L3 glia. Thus, glia are both glycolytic and oxidative
457 in the embryo and transition to being primarily oxidative in the larva. By contrast, neurons undergo
458 a clear metabolic shift from glycolytic in the embryo to oxidative in the larva, as L3 neurons
459 display high expression of most TCA enzymes, including *PDH*.

460 Both E16 glia and neurons show significant enrichment of lipogenic transcripts. The key enzyme,
461 *ACC*, and TAG biosynthetic enzymes (*FAS*, *Gpat*, *Agpat*, *Lpin*) are highly expressed. Interestingly,

462 the expression of enzymes involved in lipid break down, such as *Acyl-CoA oxidase (AOX)*, the
463 first enzyme of the beta-oxidation system, and *Carnitine O-palmitoyltransferase (CPT1* and
464 *CPT2)*, which transfers fatty acids into the mitochondrion for β -oxidation, is also high in E16
465 neurons but not glia. Thus, L3 glia continues to show elevated expression of a subset of TAG
466 biosynthetic enzymes (*FAS, Gpat, Agpat, Lpin*). Additionally, L3 glia show high expression of the
467 fatty acid breakdown pathway. Taken together, these data suggest that glia are lipogenic both in
468 embryo and larvae but activate lipolytic genes specifically in the larval stage. Neurons, on the other
469 hand, sustain a lipogenic and lipolytic state in the embryo but in the larva, they are only lipogenic.

470

471

472 **Discussion**

473 The definition of cell types makes the object of intense investigation for the potential impact on
474 medical science and in particular on regenerative medicine. We here identify the molecular
475 signatures that characterize *Drosophila* neurons, glia and hemocytes, as representative examples
476 of related and unrelated cell types. We show that the molecular identity of a cell type results from
477 the combined action of intrinsic and extrinsic cues and we establish the impact of origin, position,
478 metabolic state, function and time. We identify cell-specific as well as stage-specific signatures
479 and we show that cells maintain their identity during development while keeping their ability to
480 accommodate to the evolving needs of the organism.

481

482 **Cell types are defined by a relatively low number of specific transcripts**

483 Several technologies allow the genome-wide characterization of cells at the molecular level. Bulk
484 transcriptomes provide an adapted tool to extrapolate the most significant and specific features
485 that identify a cell type as a whole, due to the sequencing depth and to the average measurement
486 of gene expression across a population of cells. Using this approach, we show that the majority of
487 the genes are not differentially expressed between neurons, glia and hemocytes, both in the embryo
488 and in the larva. Such high number of similar genes cannot be due to common origin/function
489 because neurons and hemocytes share neither. This data nevertheless goes along with a study

490 showing that 48.8% of all Flybase genes are non-differentially expressed between all stages
491 (Daines et al., 2011). No more than 2000 genes are upregulated in each cell type, hence, a relatively
492 low number of genes is sufficient to define a cell type.

493 In average, the transcripts enriched in a specific cell type are expressed at high levels, suggesting
494 that they are under the control of signaling pathway rather than being expressed constitutively and
495 at low levels. Thus, cell types are defined by a combination of cell-specific transcript enrichment
496 and high levels of expression. Of note, even amongst the common transcripts we find a similar
497 distinction: the genes that have a house keeping function shared by all cell types are expressed at
498 lower levels than those that are associated with more specific functions. For instance, the
499 comparison of neurons and glia shows that the genes involved in DNA replication display a
500 number of reads < 300 while those related to axons (which are common to neurons and glia but
501 not to hemocytes) are expressed at higher levels (> 3000 reads).

502 In sum, cell types are defined by few genes expressed at high levels.

503

504 **Transcriptional landscapes and the impact of time**

505 The establishment of a cell type involves the stable expression of specific morphological and
506 functional features, even though cells may lose some traits or assume others over time. We hence
507 expected each cell type to group together throughout development. Instead, the developmental
508 stage represents the first factor of homogeneity, more than the cell type: embryonic hemocytes are
509 closer to embryonic glia and neurons than to larval hemocytes. Thus, although differentiated cells
510 accomplish the same function throughout the life of the organism, time affects their state.

511 Amongst the stage-specific genes, ECM related GO terms are upregulated in E16 compared to L3
512 cells. In line with the fact that the ECM is a key player in morphogenesis and organogenesis,
513 different types of embryonic cells likely express large quantities of ECM transcripts to prepare for
514 the larval life and the immense growth happening during this stage. L3 cells show an enrichment
515 of transcripts involved in DNA damage and repair. In particular, the three components of the MRN
516 complex (Rad50, Mre-11 and NBS) and Tefu, that respond to DNA DSB and are involved in
517 replication fork dynamics, telomere maintenance and even response to viral infection (Ciapponi et
518 al., 2004; Syed and Tainer, 2018). The increased expression of these genes is seemingly not due

519 to DSB since there are more DBS markers in E16 than in L3. Our results suggest that Rad50
520 functions beyond DNA repair and controls gene expression through the modulation of chromatin
521 state, in line with recent finding showing that the yeast ortholog represses the loci near its binding
522 site (Forey et al., 2021). Future studies will address whether the changes in both euchromatic and
523 heterochromatic marks observed in the mutant larval salivary glands are due to late, indirect,
524 effects or reflect a novel regulatory mechanism.

525 The RNAseq data suggest that cells are able to sense time, a process that at least in the case of
526 neurons is disconnected from the cell cycle and from cell division, as these cells do not undergo
527 mitosis. The upregulation of genes involved in histone modification and chromatin remodeling in
528 L3 suggests a stage-specific epigenetic control of the transcriptional landscape.

529 Finally, the analysis of the transcription factors at the two stages suggests that the three cell types
530 are endowed with distinct degrees of plasticity. The E16 and the L3 hemocytes share many fewer
531 transcription factors (n=25, **Sup. Table 1-i,j in red**) than those that are common to the three
532 embryonic cell types (n=129, **Sup. Table 1-a**). This behavior is also found in glia (n=17, **Sup.**
533 **Table 7-k,l in yellow**) but not in neurons (n= 124, **Sup. Table 1-m,n in blue**), which share more
534 than 50% of upregulated transcription factors at E16 and L3. The transcriptome of neurons as well
535 as the transcription factors shared at E16 and L3 suggest that neurons are a relatively stable
536 population compared to hemocytes. In the case of glia, though comparing their transcriptome at
537 E16 and L3 shows similar behavior to neurons, the fact that they share few transcription factors at
538 E16 and L3 suggests an intermediate level of stability. It is tempting to speculate that the neural-
539 immune phenotype of glial cells and the strong reliance of hemocytes on the environment could
540 account for the plastic behavior of these cell types. An increased number of glial and hemocytes
541 subtypes in larval stages could also account for such behavior. Future single cell analyses on the
542 different cell populations at different stages will help distinguishing between these possibilities.

543

544 **Hemocytes and glia, macrophages outside and within the nervous system**

545 The origin and position of neurons and glia have a significant impact on their transcriptional
546 landscapes, as shown by the large number of common molecular features. A substantial fraction
547 of the shared GO terms concern processes related to axons, synapses, and neural processes. In
548 contrast, neurons and hemocytes share many fewer transcripts (5,9% at E16, 3,8% at L3), with no

549 GO terms related to cell-specific processes (**Sup. Table 8-a,b**). The situation is different for the
550 glia/hemocyte comparison. E16 glia and hemocytes share the same very low degree of similarity
551 as that observed between neurons and hemocytes (5,7%). In addition, genes upregulated in E16
552 hemocytes do not seem to be involved in hemocyte-specific functions. This is likely due to the fact
553 that the major function of the E16 hemocytes is to secrete ECM components and to phagocytose
554 apoptotic cells, both of which are also done by glia. By contrast, L3 hemocytes and glia share
555 18,3% of enriched transcripts, some of which are related to immunity. The presence of hemocyte-
556 specific GO in the larva indicates a real developmental switch (see also Cattenoz et al., 2020) and
557 a strong specialization of the hemocytes, with more refined groups of genes being upregulated and
558 a strong immune component.

559 Glia are not just macrophages that are also able to perform neural functions. Much like microglia,
560 the vertebrate myeloid cells that invade the nervous system during development (Neumann et al.,
561 2009; Wu et al., 2015) (DePaula-Silva et al., 2019; Hickman et al., 2013; Li and Barres, 2017;
562 Ritzel et al., 2015), fly glia express very specific scavenging features compared to the immune
563 cells located outside the nervous system. Typically, *NimA* is not expressed in hemocytes but only
564 in a subset of glia. Pursuing the role of *NimA* will help deciphering the mode of action of glia in
565 the immune response, which so far has relied only on the study of Draper and NimC4, the latter
566 being only expressed in embryonic glia. *NimA* is likely not the only additional scavenger receptor
567 possibly involved in the phagocytosis of debris and apoptotic bodies. A recent study identified the
568 role of *NimB4* in apoptotic bodies phagocytosis in glia and in hemocytes in the larva (Petrigiani
569 et al., 2021), even though *NimB4* expression in hemocytes is at least 70-fold higher than in glia (our
570 data).

571 The RNAseq data highlight an intermediate neuro-immune phenotype of fly glia. We propose that
572 simple organisms display relatively few cell types with multiple potentials whereas more complex
573 organisms need a more refined division of labor accompanied by the appearance of dedicated cell
574 types, such as the microglia in vertebrates. Interestingly, the comparative analysis of the
575 transcription factor reveals the upregulation of the Nuclear factor of activated T-cells (NFAT)
576 ortholog in L3 glia and hemocytes compared to neurons (**Sup. Table 1-g in gray**). While the
577 *Drosophila* gene has not yet been explored in the context in immunity, NFAT is a member of a
578 Calcium dependent protein family that plays an important role in the immune response in lymphoid
579 and myeloid vertebrate lineages, including microglia (reviewed in Müller and Rao, 2010).

580 In sum, hemocytes become more specialized as the animal hatches. Fly glia are at the interface
581 between the nervous and the immune systems: the neural origin shared with neurons is already
582 apparent in the embryo, while the immune function shared with hemocytes becomes apparent in
583 the larva. Given the importance of the immune resident cells of the CNS in development and
584 homeostasis, fly glia represent an interesting tool to unravel the role of myeloid-like cells in
585 diseases as severe and diverse as brain tumor, neurodegeneration and autoimmune diseases.

586

587 **Cell and stage-specific metabolic states**

588 Metabolism and cell identity are closely coupled so that cells can carry out their function and meet
589 the changing needs during the life cycle. Our transcriptomic data reveal the following key
590 observations: a) all cell types show high levels of *ldh* expression in the embryo, b) they appear
591 predominantly lipogenic in the embryo and c) they switch from being glycolytic in the embryo to
592 being oxidative in the larva. While these features are shared by the three cell types across
593 development, there are stark differences in terms of glucose uptake and subsequent break down.
594 E16 hemocytes rely on *de novo* glucose and lipid biosynthesis and sustain a glycolytic state, while
595 their larval counterpart depend more on the uptake of sugars and lipids and their subsequent
596 breakdown to generate intermediates that run the TCA to sustain an oxidative state. E16 glia
597 internalize sugars, generate lipids and metabolize them via glycolytic and oxidative routes, while
598 L3 glia rely on gluconeogenesis and TAG biosynthesis and their breakdown to drive TCA and
599 maintain an oxidative state. E16 neurons mediate sugar uptake and *de novo* lipogenesis and sustain
600 a glycolytic state, whereas L3 neurons are more oxidative, which could be driven by *de novo* sugar
601 synthesis and its breakdown to sustain TCA. L3 neurons also show reliance on TAG biosynthesis,
602 which is perhaps critical for generating lipids essential for synaptic vesicle release. We speculate
603 that in E16 neurons lipid synthesis and catabolism may function as a potential energy source while
604 in E16 glia, lipid synthesis may be relevant for functions like secretory vesicular based intercellular
605 communication.

606 In sum, embryonic cells are capable of intracellular synthesis of sugars and lipids and are also
607 involved in internalizing extracellular metabolites. Later on, neurons and glia indulge in active
608 uptake of metabolites and also continue to drive intracellular synthesis of lipids and sugars. Thus,
609 both extracellular and intra-cellular resources drive the TCA in these tissues. By contrast, L3

610 hemocytes appear more reliant on extracellular nutrients to drive their internal lipid oxidation and
611 TCA catabolic pathways. Such reliance on extracellular metabolites as the predominant source
612 makes hemocytes competent sensors of systemic metabolites and any fluctuation in their levels.

613 The analyses of the metabolic pathways revealed an unexpected aspect of stage-specificity: the
614 utilization of distinct paralogs in embryos and larvae. The fatty acid metabolic enzyme, *Fatty acid*
615 *synthase 1* (*FASN1*, *FBgn0027571*), *CG3523*, is expressed in larvae, while its paralog *CG3524*
616 (*FASN2*, *FBgn0042627*), was detected in embryos. The enzyme of lipid breakdown pathway,
617 *CG17544* (*FBgn0032775*) with predicted acyl-CoA oxidase activity, is expressed in larvae, while
618 its paralog *CG9709*, *acyl-Coenzyme A oxidase at 57D distal* (*Acox57D-d*, *FBgn0034629*), in
619 embryos. Additionally, we identified genes that are not referred to as paralogs in Flybase, but share
620 functional similarity, showing a similar trend: *CG6650* (*FBgn0036402*), predicted to have ADP-
621 glucokinase activity, is expressed in larvae, while the glycolytic enzyme *CG8094*, *Hexokinase C*
622 (*Hex-C*, *FBgn0001187*), in embryos. For the TCA enzymes, *CG7024* (*FBgn0029722*), predicted
623 to have pyruvate dehydrogenase activity, is expressed in larvae while *CG11876*, which is *Pyruvate*
624 *dehydrogenase E1 beta subunit* (*Pdhb*, *FBgn0039635*), in embryos. The expression of *CG12233*,
625 *Idh3a* (*FBgn0027291*), was detected in larvae, while *CG32026* (*FBgn0052026*), predicted with
626 isocitrate dehydrogenase activity, was noticed in embryos. Finally, the expression of *Ogdh*
627 (*Fbgn0010352*), *CG11661*, was seen in embryos, while *CG5214* (*FBgn0037891*), predicted to be
628 part of the oxoglutarate dehydrogenase complex, in larvae. The enzymes with predicted acyl-CoA
629 activity are tandemly duplicated, but the vast majority of the other genes are distantly located or
630 reside on different chromosomes, calling for independent transcriptional regulation and/or stage-
631 specific differences in their enzymatic activities. Future studies will establish the overall relevance
632 of paralog switching on metabolic state transitions, but such temporal specificity provides an
633 unexpected level of control that adds to the post transcriptional modifications of those enzymes.

634

635 The unbiased identification of common and specific transcriptional pathways allows a better
636 understanding of the molecular processes associated to cell type identity. This genome-wide study
637 has identified the transcriptional features of three types of related and unrelated cells, including
638 their metabolic states and their transcriptional regulators. It has revealed novel cell-specific and
639 stage-specific players including NimA and Rad50, respectively. It has highlighted time as a novel

640 parameter defining a differentiated cell, paving the way to understand the role of high-order
641 chromatin organization in the process. Altogether, this comparative high throughput analysis
642 allows a better understanding of the mechanisms defining cell type identity, a central issue in
643 biology and medical science.

644

645

646 **Materials and methods**

647 **Fly strains and genetics**

648 Flies were raised on standard medium at 25°C. Fly strains used are: *srp(hemo)Gal4/+; UAS-*
649 *RFP/+* obtained upon crossing *srp(hemo)Gal4* (gift from K. Brückner) (Brückner et al., 2004) and
650 *UAS-RFP* flies (RRID:BDSC_8547), *HmlA-RFP/+* (Makhijani et al., 2011), *srp(hemo)3xmcherry*
651 (gift from D. Siekhaus, (Gyoergy et al., 2018), *repo-nRFP 43.1* on the 3rd chromosome (Laneve
652 et al., 2013), *elav-nRFP 28.2* insertion on the 3rd chromosome *nSybGal4/+; UAS-RFP/+* obtained
653 upon crossing *nSybGal4/+* (RRID:BDSC_51635) with *UAS-RFP/+*, *rad50Δ5.1* (Ciapponi et al.,
654 2004) and *Oregon-R* strain was used as the control (referred to as *WT* in text). *NimAGal4* null
655 mutant was generated by CRISPR-mediated mutagenesis and performed by WellGenetics Inc.
656 using modified methods from (Kondo and Ueda, 2013). In brief, the upstream gRNA sequence
657 ACTGCTCCTCCTGCTTGCAA[TGG] and downstream gRNA sequence
658 GTGCCCTTCTAACATATAACC[AGG] were cloned into U6 promote plasmid(s). Cassette *Gal4-*
659 *3xP3-RFP*, which contains ribosome binding sequence (RBS), Gal4, SV40 polyA terminator and
660 a floxed 3xP3-RFP, and two homology arms were cloned into pUC57-Kan as donor template for
661 repair. *NimA*-targeting gRNAs and hs-Cas9 were supplied in DNA plasmids, together with donor
662 plasmid for microinjection into embryos of control strain *w[1118]*. F1 flies carrying selection
663 marker of 3xP3-RFP were further validated by genomic PCR and sequencing. CRISPR generates
664 a 4,884-bp deletion removing the entire *NimA* CDS that is replaced by the cassette *Gal4-3xP3-*
665 *RFP. NimAGal4; UAS-eGFP* animals were obtained upon crossing *NimAGal4* with *UAS-eGFP*.

666 **FACS sorting of hemocytes, glia and neurons**

667 Embryonic cells were isolated as follows. Staged egg laying was carried out to produce E16
668 embryos. *srp(hemo)Gal4/+; UAS-RFP/+*, *repo-nRFP*, *elav-nRFP* and *Oregon-R* strains were
669 amplified, staged lays were done on yeast apple juice agar at 25°C. After a pre-lay period of 30
670 minutes, the agar plates and yeast were replaced with fresh plates and flies were left to lay for 3
671 hours at 25°C. Agar plates were removed and kept at 25°C, embryos were collected 11 hours and
672 40 minutes AEL, when they had reached stage 16. Embryos were then isolated from the medium
673 and washed on a 100 µm mesh. The collected embryos were transferred into a cold solution of
674 phosphate-buffered saline (PBS) in a Dounce homogenizer on ice. The embryos were dissociated
675 using the large clearance pestle then the small clearance pestle, then filtered (70 µm filter).

676 L3 neurons, glia and hemocytes were purified from *elav-nRFP*, *repo-nRFP* and *HmlΔ-RFP/+*
677 larvae, respectively. Staged lays of 3 hours were carried out at 25°C and wandering larvae were
678 collected 108–117 hours AEL. L3 hemocytes were isolated as mentioned in (Cattenoz et al., 2020).
679 Larvae were bled in cold PBS containing PTU (Sigma-Aldrich P7629) to prevent hemocyte
680 melanization (Lerner and Fitzpatrick, 1950), and filtered (70 µm filter). For neurons and glia
681 isolation, larval brains were dissected in cold PBS on ice and transferred to tubes containing 0.5
682 µg of collagenase IV (Gibco, Invitrogen) in 220 µL of PBS. Brains were incubated at 37°C on a
683 Thermomixer with shaking at 500 rpm for 20 minutes. Then the brains were dissociated by
684 pipetting up and down with 10-gauge needles and syringes and filtered through a 70 µm filter.

685 Cells were sorted using FACS Aria II (BD Biosciences) at 4°C in three independent biological
686 replicates for each genotype. Live cells were first selected based on the forward scatter and side
687 scatter and only single cells were sorted according to the RFP signal. *Oregon-R* cells were used as
688 a negative control to set the gate and sort the RFP positive cells (**Sup. Figure 3-n**), which were
689 collected directly in TRI reagent (MRC) for RNA extraction. Around 100 000 cells were sorted
690 for each replicate. The purity of the sorted populations was assessed by carrying out a post-sort
691 step. The FACS sorter was set up to produce cell pools displaying at least 80% of purity on the
692 post-sort analysis.

693 **RNA extraction and sequencing**

694 Sorted cells were homogenized in TRI reagent (MRC) for 5 minutes at room temperature (RT) to
695 ensure complete dissociation of nucleoprotein complexes. 0.2 mL of chloroform was added to each
696 sample followed by centrifugation at 12,000 g for 15 minutes at 4°C. The upper aqueous phase
697 containing the RNA was collected and transferred to a new tube. 0.5 mL of 2-propanol were added,
698 and the samples were incubated for 10 minutes at RT. The samples were then centrifuged at 12,000
699 g for 10 minutes to precipitate the RNA. The RNA pellet was then washed with 1 mL of 75%
700 ethanol then precipitated again at 7,500 g for 5 minutes and air dried. 20 µL of RNase-free water
701 was added to each sample before incubation at 55°C for 15 minutes. Single-end polyA+ RNA-Seq
702 (mRNA-seq) libraries were prepared using the SMARTer (Takara) Low input RNA kit for
703 Illumina sequencing. All samples were sequenced in 50-length Single-Read.

704 **Data analysis**

705 The data analysis was done using the GalaxEast platform, the Galaxy instance of east of France
706 (<http://www.galaxeast.fr/>). First, FastQC (Babraham Bioinformatics) was used on the raw FastQ
707 files to generate summary statistics and assess the quality of the data. The raw files were then
708 converted to FastQ Sanger using FastQ Groomer for downstream analysis. Data were then mapped
709 to the *Drosophila melanogaster* August 2014 Dm6 (BDGF release 6 + ISO1 MT/Dm6) reference
710 genome using TopHat (Trapnell et al., 2009). The number of reads per annotated gene were
711 counted using Htseq-Count (Anders et al., 2015) and the comparison and normalization of the data
712 between the different cell types was done using Deseq2 (Anders and Huber, 2010). To compare
713 E16 stage to L3 using Deseq2, we considered all cell types deriving from E16 as replicates and
714 compared to the cell deriving from L3 that were also considered replicates. Gene ontology studies
715 were done using the Panther classification system version 16 (<http://pantherdb.org/>) (Mi et al.,
716 2010; Thomas et al., 2003) for the identification of biological processes, Bonferroni correction
717 was used for all analyses. Genes with a number of reads > 15 were considered significantly
718 expressed. For the study of upregulated genes, only genes with fold change >1.5, adjusted p-value
719 <0.05 and expression level >15 were taken into account. Genes with number of reads > 15 and
720 adjusted p-value of fold change > 0.05 were considered non-differentially expressed. Amongst
721 these, genes with number of reads > 15 and fold change < 1.5 were considered commonly
722 expressed. For the study of scavenger receptors only genes with expression level > 500, adjusted
723 p-value < 0.05 and fold change > 10 were considered as specific to either glia or hemocytes. The

724 Venn diagrams were generated using Venny 2.1 (<https://bioinfogp.cnb.csic.es/tools/venny/>)
725 (Oliveros, J.C. (2007-2015)) and data representation was done using Microsoft Excel. The
726 heatmaps representing the log₁₀ of the absolute expression levels in glia, neurons and hemocytes
727 at E16 and L3 or representing the z-score were plotted in R (version 3.4.0) (R CoreTeam) using
728 pheatmap (version 0.2) package (Kolde and Kolde, 2015).

729 **Quantitative PCR**

730 For the comparison between the expression levels of scavenger receptors in hemocytes and glia,
731 cells were isolated from *srp(hemo)-3xmCherry* and *repo-nRFP* wandering L3, respectively. FACS
732 sorting and RNA extraction were as described above. The extracted RNA was then treated with
733 DNase I recombinant RNase free (Roche) and the reverse transcription was done using the Super-
734 Script IV (Invitrogen) with random primers. The cycle program used for the reverse transcription
735 is 65°C for 10 minutes, 55°C for 20 minutes, 80°C for 10 minutes. The qPCR was done using
736 SYBR Green I Master (Roche). Actin5C (Act5C) and Ribosomal protein 49 (RP49) were used to
737 normalize the data. The primers are listed in **Sup. Table 9**. The p-values and statistical test used
738 are indicated in the figure legends.

739 For the quantitative PCR on larval and embryonic tissues, whole larvae and whole embryos were
740 crushed in cold PBS using a Dounce homogenizer, extracts were filtered first through a cell strainer
741 of 100 µm and RNA extraction, reverse transcription and qPCR were done as described.

742 **Histone extraction and western blot**

743 Histones from E16 and L3 *Oregon-R* were extracted as described in Abcam histone extraction
744 protocol. Histone extracts were separated by 15% SDS-PAGE, transferred onto nitrocellulose
745 membrane and probed with the primary antibodies mouse anti- H2A.v-P (1:5000) recognizing the
746 *Drosophila* Histone 2A gamma variant, phosphorylated H2A.v-P (Developmental Studies
747 Hybridoma Bank (DSHB AB_2618077, Lake et al., 2013) and rabbit anti-H3 (1:5000) (Abcam
748 ab1791) for normalization. Signal was detected with Pierce ECL western blotting substrate
749 (Thermo Fisher Scientific, Waltham, MA) using HRP-conjugated secondary antibodies (1:10000,
750 Jackson).

751 **Immunolabeling of central nervous system**

752 *NimAGal4; UAS-eGFP* embryos were dechorionated in bleach for 5 minutes and rinsed in water.
753 They were then fixed in 50% heptane/50% paraformaldehyde (PFA) for 25 minutes on a shaker at
754 RT and devitellinized in methanol/heptane for 1 minute. Embryos were then rinsed in methanol
755 followed by PTX (0.3% Triton X-100 in PBS 1x) and incubated in blocking reagent (Roche) for 1
756 hour at RT. They were incubated with primary antibodies mouse anti-Repo 1:20 (DSHB #8D12),
757 chicken anti-GFP 1:500 (Abcam ab13970) and rabbit anti-Nazgul 1:100 (gift From B. Altenhein)
758 in blocking reagent overnight (O/N) at 4°C, then washed in PTX 3 × 10 minutes at RT and
759 incubated with secondary antibodies 1:400 (Cy3 anti-mouse and FITC anti-chicken (Jackson)) for
760 1 hour at RT and washed 3x 10 minutes at PTX. The embryos were mounted in Vectashield
761 (Vector #H-1000) and analyzed by confocal microscopy (Leica spinning disk). For L3 larvae and
762 adults, CNSs from *NimAGal4 ; UAS-eGFP* were dissected in cold PBS 1x and transferred into
763 wells containing 4% PFA and fixed for 2 hours at RT. They were then washed in PTX for 1 hour
764 and incubated in blocking reagent for 1 hour at RT. CNSs were then incubated with primary
765 antibodies mouse anti-Repo, chicken anti-GFP and rabbit anti-Nazgul O/N at 4°C, then washed in
766 PTX 3x 10 minutes and incubated with secondary antibodies (Cy5 anti-mouse, Cy3 anti-rabbit
767 (Jackson)) for 1 hour at RT. After washing in PTX, CNSs were mounted in Vectashield imaged
768 using Leica spinning disk microscope and analyzed using Fiji (National Institute of Mental Health,
769 Bethesda, Maryland, USA, (Schindelin et al., 2012).

770 **Immunolabeling of polytene chromosomes**

771 Salivary glands were dissected from L3 larvae in NaCl 0.7% / NP-40 0.1% and fixed for 2 minutes
772 in 3.7% formaldehyde/ 0.7% NaCl/ 1% Triton-X. Before squashing, the glands were transferred
773 in 2% formaldehyde/ 45% acetic acid and incubated for 2 minutes on a siliconized coverslip.
774 During incubation, salivary glands were fragmented. A poly-L-lysine slide was inverted on top of
775 the coverslip and salivary glands were squashed. The chromosome spreads were examined by
776 phase microscopy and frozen in liquid nitrogen. The coverslip was flicked off with a razor blade
777 and slides were incubated in cold PBS 2× for 15 min. The slides were transferred to a blocking
778 solution (3% BSA, 2% NP-40, 0.2% Tween-20, 10% non-fat dry milk in PBS) and blocked for 1
779 hour at RT. After blocking, slides were incubated with primary antibody in blocking solution
780 overnight at 4°C in a humid chamber. Slides were then rinsed in PBS and washed 2x 15 minutes
781 in PBS 1x /0.2% Tween/0.1% NP-40. After washing, slides were rinsed in PBS and incubated with

782 secondary antibody in blocking solution 2 hours at RT in a humid chamber. Slides were then rinsed
783 in PBS and washed 2x 15 minutes in PBS1x /0.2% Tween/0.1% NP-40. Finally, slides were
784 drained and mounted with Vectashield medium H-1200 with DAPI to stain DNA. The primary
785 antibodies used for immunolabeling were: rabbit anti-H3K4me3 1:400 (Abcam ab8580); mouse
786 anti-H3K9me2 1:20 (Abcam ab1220); rabbit anti-H3K9me3 1:100 (Abcam ab8898); rabbit anti-
787 H3K27me3 1:400 (Cell Signaling C36B11); mouse anti-H2A.v-P 1:20 (DSHB AB_2618077). The
788 secondary antibodies were: FITC-conjugated goat anti-rabbit 1:50 (Jackson) and Rhodamine
789 conjugated goat anti-mouse 1:50 (Jackson). Polytene chromosomes preparations were analyzed on
790 a fluorescence microscope (Zeiss Apotome) and images acquisition performed using the Zen Pro
791 software (Zeiss). Polytene chromosome fluorescence intensity and bands plot profile analysis were
792 performed using Fiji software (National Institute of Mental Health, Bethesda, Maryland, USA).

793 ***In situ* hybridization**

794 We used a RNAscope® probe and reagent kit. Larval CNSs from *Oregon R* were dissected in cold
795 PBS 1x and fixed in 4% PFA O/N at 4°C then washed 3x 10 minutes in PBST (PBS 1x/ 0.1%
796 Tween-20) followed by one wash in PBS 1x. Samples were then incubated in RNAscope reagents
797 as in protease III at 40°C for 10 minutes and washed in 3x 2 minutes in PBS 1x. The probe
798 targeting *NimA* was then added in addition to the positive control probe and samples were
799 incubated O/N at 40°C. The amplifiers and detection solution in the RNAscope reagent kit were
800 added sequentially for hybridization signal amplification for the indicated time. CNSs were then
801 mounted in Vectashield, imaged using Leica spinning disk microscope and analyzed using Fiji.

802 **Metabolic pathway analysis**

803 All data points considered for our analysis has a read count of more than ten in at least one of the
804 samples from the tissue under consideration. Differential expression analysis between the L3 and
805 E16 developmental stages was done for each tissue using DESeq2. Genes associated with
806 metabolic pathways considered in this study, were retrieved from the KEGG database (Kanehisa
807 and Goto, 2000, <http://www.genome.jp/>). The heatmaps representing the z-score of the metabolic
808 genes in glia, neurons and hemocytes at E16 and L3 were plotted using R (version 3.4.0) (R
809 CoreTeam). For genes with paralogs, the paralogs show $\log_2FC(L3/E16)$ greater or lesser than
810 zero, the paralog with the highest fold-change has been plotted. For genes whose paralogs have
811 $\log_2FC(L3/E16)$ that is either positive or negative, the paralogs with the highest and lowest fold-

812 change have been plotted. The details of all the genes (including all the paralogs) as well as genes
813 included in the figures are present in **Sup. Table 7**.

814

815

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832

833

834 **Author contribution**

835 Conceptualization, RS and AG ; Methodology, RS, AG, PBC, RS, AG, PBC, AP, MM, LC, TM
836 and NH. Investigation, RS, AG, PBC, AP, MM, LC, TM and NH. Writing—Original Draft, RS
837 and AG, Writing—Review and Editing, RS, AG and PBC; Funding Acquisition, AG, LC and TM;
838 Resources, AG; Supervision, AG and PBC.

839

840 **Conflict of interest**

841 The authors declare that they have no conflict of interest.

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843

844 **References**

845 Abbott, N.J. (2005). Dynamics of CNS barriers: evolution, differentiation, and modulation. *Cell. Mol. Neurobiol.* *25*, 5–23.

847 Anders, S., and Huber, W. (2010). Differential expression analysis for sequence count data. *Genome Biol.* *11*.

849 Anders, S., Pyl, P.T., and Huber, W. (2015). HTSeq--a Python framework to work with high-throughput sequencing data. *Bioinformatics* *31*, 166–169.

851 Aranda, S., Mas, G., and Di Croce, L. (2015). Regulation of gene transcription by Polycomb proteins. *Sci. Adv.* *1*, e1500737.

853 Awasaki, T., and Ito, K. (2004). Engulfing action of glial cells is required for programmed axon pruning during *Drosophila* metamorphosis. *Curr. Biol.* *14*, 668–677.

855 Barres, B.A. (2008). The mystery and magic of glia: a perspective on their roles in health and disease. *Neuron* *60*, 430–440.

857 Basenko, E.Y., Sasaki, T., Ji, L., Prybol, C.J., Burckhardt, R.M., Schmitz, R.J., and Lewis, Z.A. (2015). Genome-wide redistribution of H3K27me3 is linked to genotoxic stress and defective growth. *Proc. Natl. Acad. Sci. U. S. A.* *112*, E6339-48.

860 Bittern, J., Pogodalla, N., Ohm, H., Brüser, L., Kottmeier, R., Schirmeier, S., and Klämbt, C. (2021). Neuron-glia interaction in the *Drosophila* nervous system. *Dev. Neurobiol.* *81*, 438–452.

862 Bosso, G., Cipressa, F., Moroni, M.L., Pennisi, R., Albanesi, J., Brandi, V., Cugusi, S., Renda, F., Ciapponi, L., Polticelli, F., et al. (2019). NBS1 interacts with HP1 to ensure genome integrity. *Cell Death Dis.* *10*, 951.

865 Brink, D.L., Gilbert, M., Xie, X., Petley-Ragan, L., and Auld, V.J. (2012). Glial processes at the *Drosophila* larval neuromuscular junction match synaptic growth. *PLoS One* *7*, e37876.

867 Brown, N.H. (2011). Extracellular matrix in development: insights from mechanisms conserved between invertebrates and vertebrates. *Cold Spring Harb. Perspect. Biol.* *3*.

869 Brückner, K., Kockel, L., Duchek, P., Luque, C.M., Rørth, P., and Perrimon, N. (2004). The PDGF/VEGF receptor controls blood cell survival in *Drosophila*. *Dev. Cell* *7*, 73–84.

871 Callebaut, I., Mignotte, V., Souchet, M., and Mornon, J.P. (2003). EMI domains are widespread and reveal the probable orthologs of the *Caenorhabditis elegans* CED-1 protein. *Biochem. Biophys. Res. Commun.* *300*, 619–623.

874 Cattenoz, P.B., Sakr, R., Pavlidaki, A., Delaporte, C., Riba, A., Molina, N., Hariharan, N., Mukherjee, T., and Giangrande, A. (2020). Temporal specificity and heterogeneity of *Drosophila* immune cells. *EMBO J.* e104486.

877 Ciapponi, L., Cenci, G., Ducau, J., Flores, C., Johnson-Schlitz, D., Gorski, M.M., Engels, W.R., and Gatti,

878 M. (2004). The *Drosophila* Mre11/Rad50 complex is required to prevent both telomeric fusion and
879 chromosome breakage. *Curr. Biol.* *14*, 1360–1366.

880 Ciapponi, L., Cenci, G., and Gatti, M. (2006). The *Drosophila* Nbs protein functions in multiple pathways
881 for the maintenance of genome stability. *Genetics* *173*, 1447–1454.

882 Conway, E., Healy, E., and Bracken, A.P. (2015). PRC2 mediated H3K27 methylations in cellular identity
883 and cancer. *Curr. Opin. Cell Biol.* *37*, 42–48.

884 Cornman, R.S. (2009). Molecular evolution of *Drosophila* cuticular protein genes. *PLoS One* *4*, e8345.

885 Daines, B., Wang, H., Wang, L., Li, Y., Han, Y., Emmert, D., Gelbart, W., Wang, X., Li, W., Gibbs, R., et
886 al. (2011). The *Drosophila melanogaster* transcriptome by paired-end RNA sequencing. *Genome Res.* *21*,
887 315–324.

888 Davie, K., Janssens, J., Koldere, D., De Waegeneer, M., Pech, U., Kreft, Ł., Aibar, S., Makhzami, S.,
889 Christiaens, V., Bravo González-Blas, C., et al. (2018). A Single-Cell Transcriptome Atlas of the Aging
890 *Drosophila* Brain. *Cell* *174*, 982–998.e20.

891 Deliu, L.P., Ghosh, A., and Grewal, S.S. (2017). Investigation of protein synthesis in *Drosophila* larvae
892 using puromycin labelling. *Biol. Open* *6*, 1229–1234.

893 DePaula-Silva, A.B., Gorbea, C., Doty, D.J., Libbey, J.E., Sanchez, J.M.S., Hanak, T.J., Cazalla, D., and
894 Fujinami, R.S. (2019). Differential transcriptional profiles identify microglial- and macrophage-specific
895 gene markers expressed during virus-induced neuroinflammation. *J. Neuroinflammation* *16*, 1–20.

896 Ebens, A.J., Garren, H., Cheyette, B.N., and Zipursky, S.L. (1993). The *Drosophila* anachronism locus: a
897 glycoprotein secreted by glia inhibits neuroblast proliferation. *Cell* *74*, 15–27.

898 Forey, R., Barthe, A., Tittel-Elmer, M., Wery, M., Barrault, M.-B., Ducrot, C., Seeber, A., Krietenstein, N.,
899 Szachnowski, U., Skrzypczak, M., et al. (2021). A Role for the Mre11-Rad50-Xrs2 Complex in Gene
900 Expression and Chromosome Organization. *Mol. Cell* *81*, 183–197.e6.

901 Franc, N.C., Dimarcq, J.L., Lagueux, M., Hoffmann, J., and Ezekowitz, R.A. (1996). Croquemort, a novel
902 *Drosophila* hemocyte/macrophage receptor that recognizes apoptotic cells. *Immunity* *4*, 431–443.

903 Franc, N.C., Heitzler, P., Ezekowitz, R.A., and White, K. (1999). Requirement for croquemort in
904 phagocytosis of apoptotic cells in *Drosophila*. *Science* *284*, 1991–1994.

905 Garces, A., and Thor, S. (2006). Specification of *Drosophila* aCC motoneuron identity by a genetic cascade
906 involving even-skipped, grain and *zfh1*. *Development* *133*, 1445–1455.

907 Gatei, M., Kijas, A.W., Biard, D., Dörk, T., and Lavin, M.F. (2014). RAD50 phosphorylation promotes
908 ATR downstream signaling and DNA restart following replication stress. *Hum. Mol. Genet.* *23*, 4232–
909 4248.

910 Gyoergy, A., Roblek, M., Ratheesh, A., Valoskova, K., Belyaeva, V., Wachner, S., Matsubayashi, Y.,
911 Sánchez-Sánchez, B.J., Stramer, B., and Siekhaus, D.E. (2018). Tools Allowing Independent Visualization
912 and Genetic Manipulation of *Drosophila melanogaster* Macrophages and Surrounding Tissues. *G3*
913 (Bethesda). *8*, 845–857.

914 Hickman, S.E., Kingery, N.D., Ohsumi, T.K., Borowsky, M.L., Wang, L.C., Means, T.K., and El Khoury,
915 J. (2013). The microglial sensome revealed by direct RNA sequencing. *Nat. Neurosci.* *2013* *1612* *16*, 1896–
916 1905.

917 Hildebrandt, A., Pflanz, R., Behr, M., Tarp, T., Riedel, D., and Schuh, R. (2015). Bark beetle controls
918 epithelial morphogenesis by septate junction maturation in *Drosophila*. *Dev. Biol.* *400*, 237–247.

919 James, T.C., and Elgin, S.C. (1986). Identification of a nonhistone chromosomal protein associated with
920 heterochromatin in *Drosophila melanogaster* and its gene. *Mol. Cell. Biol.* *6*, 3862–3872.

921 James, T.C., Eissenberg, J.C., Craig, C., Dietrich, V., Hobson, A., and Elgin, S.C. (1989). Distribution
922 patterns of HP1, a heterochromatin-associated nonhistone chromosomal protein of *Drosophila*. *Eur. J. Cell*
923 *Biol.* *50*, 170–180.

924 Kanehisa, M., and Goto, S. (2000). KEGG: kyoto encyclopedia of genes and genomes. *Nucleic Acids Res.*
925 *28*, 27–30.

926 Karouzou, M. V, Spyropoulos, Y., Iconomidou, V.A., Cornman, R.S., Hamodrakas, S.J., and Willis, J.H.
927 (2007). *Drosophila* cuticular proteins with the R&R Consensus: annotation and classification with a new
928 tool for discriminating RR-1 and RR-2 sequences. *Insect Biochem. Mol. Biol.* *37*, 754–760.

929 Kocks, C., Cho, J.H., Nehme, N., Ulvila, J., Pearson, A.M., Meister, M., Strom, C., Conto, S.L., Hetru, C.,
930 Stuart, L.M., et al. (2005). Eater, a transmembrane protein mediating phagocytosis of bacterial pathogens
931 in *Drosophila*. *Cell* *123*, 335–346.

932 Kondo, S., and Ueda, R. (2013). Highly improved gene targeting by germline-specific Cas9 expression in
933 *Drosophila*. *Genetics* *195*, 715–721.

934 Kurant, E., Axelrod, S., Leaman, D., and Gaul, U. (2008). Six-microns-under acts upstream of Draper in
935 the glial phagocytosis of apoptotic neurons. *Cell* *133*, 498–509.

936 Kurucz, E., Márkus, R., Zsámboki, J., Folkl-Medzihradzky, K., Darula, Z., Vilmos, P., Udvardy, A.,
937 Krausz, I., Lukacsovich, T., Gateff, E., et al. (2007). Nimrod, a putative phagocytosis receptor with EGF
938 repeats in *Drosophila* plasmatocytes. *Curr. Biol.* *17*, 649–654.

939 Lake, C.M., Holsclaw, J.K., Bellendir, S.P., Sekelsky, J., and Hawley, R.S. (2013). The development of a
940 monoclonal antibody recognizing the *Drosophila melanogaster* phosphorylated histone H2A variant (γ -
941 H2AV). *G3 (Bethesda)*. *3*, 1539–1543.

942 Landgraf, M., and Thor, S. (2006). Development of *Drosophila* motoneurons: specification and
943 morphology. *Semin. Cell Dev. Biol.* *17*, 3–11.

944 Laneve, P., Delaporte, C., Trebuchet, G., Komonyi, O., Flici, H., Popkova, A., D’Agostino, G., Tagliani, F.,
945 Kerekes, I., and Giangrande, A. (2013). The Gcm/Glide molecular and cellular pathway: new actors and
946 new lineages. *Dev. Biol.* *375*, 65–78.

947 Lebestky, T., Chang, T., Hartenstein, V., and Banerjee, U. (2000). Specification of *Drosophila*
948 hematopoietic lineage by conserved transcription factors. *Science* *288*, 146–149.

949 Lemaitre, B., Nicolas, E., Michaut, L., Reichhart, J.M., and Hoffmann, J.A. (1996). The dorsoventral
950 regulatory gene cassette *spätzle/Toll/cactus* controls the potent antifungal response in *Drosophila* adults.
951 *Cell* *86*, 973–983.

952 LERNER, A.B., and FITZPATRICK, T.B. (1950). Biochemistry of melanin formation. *Physiol. Rev.* *30*,
953 91–126.

954 Li, Q., and Barres, B.A. (2017). Microglia and macrophages in brain homeostasis and disease. *Nat. Rev.*
955 *Immunol.* *2017* *184* *18*, 225–242.

956 Limmer, S., Weiler, A., Volkenhoff, A., Babatz, F., and Klambt, C. (2014). The *Drosophila* blood-brain
957 barrier: development and function of a glial endothelium. *Front. Neurosci.* *8*, 365.

958 MacDonald, J.M., Beach, M.G., Porgiglia, E., Sheehan, A.E., Watts, R.J., and Freeman, M.R. (2006). The
959 *Drosophila* cell corpse engulfment receptor Draper mediates glial clearance of severed axons. *Neuron* *50*,
960 869–881.

961 Makhijani, K., Alexander, B., Tanaka, T., Rulifson, E., and Brückner, K. (2011). The peripheral nervous
962 system supports blood cell homing and survival in the *Drosophila* larva. *Development* *138*, 5379–5391.

963 Manaka, J., Kuraishi, T., Shiratsuchi, A., Nakai, Y., Higashida, H., Henson, P., and Nakanishi, Y. (2004).

964 Draper-mediated and phosphatidylserine-independent phagocytosis of apoptotic cells by *Drosophila*
965 hemocytes/macrophages. *J. Biol. Chem.* 279, 48466–48476.

966 Mangahas, P.M., and Zhou, Z. (2005). Clearance of apoptotic cells in *Caenorhabditis elegans*. *Semin. Cell*
967 *Dev. Biol.* 16, 295–306.

968 Martinek, N., Shahab, J., Saathoff, M., and Ringuette, M. (2008). Haemocyte-derived SPARC is required
969 for collagen-IV-dependent stability of basal laminae in *Drosophila* embryos. *J. Cell Sci.* 121, 1671–1680.

970 Melcarne, C., Lemaitre, B., and Kurant, E. (2019). Phagocytosis in *Drosophila*: From molecules and cellular
971 machinery to physiology. *Insect Biochem. Mol. Biol.* 109, 1–12.

972 Mi, H., Dong, Q., Muruganujan, A., Gaudet, P., Lewis, S., and Thomas, P.D. (2010). PANTHER version
973 7: improved phylogenetic trees, orthologs and collaboration with the Gene Ontology Consortium. *Nucleic*
974 *Acids Res.* 38.

975 Naba, A., Clauser, K.R., Ding, H., Whittaker, C.A., Carr, S.A., and Hynes, R.O. (2016). The extracellular
976 matrix: Tools and insights for the “omics” era. *Matrix Biol.* 49, 10–24.

977 Neumann, H., Kotter, M.R., and Franklin, R.J.M. (2009). Debris clearance by microglia: an essential link
978 between degeneration and regeneration. *Brain* 132, 288–295.

979 O’Hagan, H.M., Mohammad, H.P., and Baylin, S.B. (2008). Double strand breaks can initiate gene
980 silencing and SIRT1-dependent onset of DNA methylation in an exogenous promoter CpG island. *PLoS*
981 *Genet.* 4, e1000155.

982 O’Hagan, H.M., Wang, W., Sen, S., Destefano Shields, C., Lee, S.S., Zhang, Y.W., Clements, E.G., Cai,
983 Y., Van Neste, L., Easwaran, H., et al. (2011). Oxidative damage targets complexes containing DNA
984 methyltransferases, SIRT1, and polycomb members to promoter CpG Islands. *Cancer Cell* 20, 606–619.

985 Ou, J., He, Y., Xiao, X., Yu, T.-M., Chen, C., Gao, Z., and Ho, M.S. (2014). Glial cells in neuronal
986 development: recent advances and insights from *Drosophila melanogaster*. *Neurosci. Bull.* 30, 584–594.

987 Parker, R.J., and Auld, V.J. (2006). Roles of glia in the *Drosophila* nervous system. *Semin. Cell Dev. Biol.*
988 17, 66–77.

989 Pearson, A., Lux, A., and Krieger, M. (1995). Expression cloning of dSR-CI, a class C macrophage-specific
990 scavenger receptor from *Drosophila melanogaster*. *Proc. Natl. Acad. Sci. U. S. A.* 92, 4056–4060.

991 Peco, E., Davla, S., Camp, D., Stacey, S.M., Landgraf, M., and van Meyel, D.J. (2016). *Drosophila*
992 astrocytes cover specific territories of the CNS neuropil and are instructed to differentiate by Prospero, a
993 key effector of Notch. *Development* 143, 1170–1181.

994 Pereanu, W., Shy, D., and Hartenstein, V. (2005). Morphogenesis and proliferation of the larval brain glia
995 in *Drosophila*. *Dev. Biol.* 283, 191–203.

996 Peters, A.H.F.M., Kubicek, S., Mechtler, K., O’Sullivan, R.J., Derijck, A.A.H.A., Perez-Burgos, L.,
997 Kohlmaier, A., Opravil, S., Tachibana, M., Shinkai, Y., et al. (2003). Partitioning and plasticity of
998 repressive histone methylation states in mammalian chromatin. *Mol. Cell* 12, 1577–1589.

999 Qin, X., Ahn, S., Speed, T.P., and Rubin, G.M. (2007). Global analyses of mRNA translational control
1000 during early *Drosophila* embryogenesis. *Genome Biol.* 8, 1–18.

1001 Rämets, M., Pearson, A., Manfrulli, P., Li, X., Koziel, H., Göbel, V., Chung, E., Krieger, M., and
1002 Ezekowitz, R.A. (2001). *Drosophila* scavenger receptor CI is a pattern recognition receptor for bacteria.
1003 *Immunity* 15, 1027–1038.

1004 Rämets, M., Manfrulli, P., Pearson, A., Mathey-Prevot, B., and Ezekowitz, R.A.B. (2002). Functional
1005 genomic analysis of phagocytosis and identification of a *Drosophila* receptor for *E. coli*. *Nature* 416, 644–
1006 648.

1007 Rea, S., Eisenhaber, F., O'Carroll, D., Strahl, B.D., Sun, Z.W., Schmid, M., Opravil, S., Mechtler, K.,
1008 Ponting, C.P., Allis, C.D., et al. (2000). Regulation of chromatin structure by site-specific histone H3
1009 methyltransferases. *Nature* 406, 593–599.

1010 Redon, C., Pilch, D., Rogakou, E., Sedelnikova, O., Newrock, K., and Bonner, W. (2002). Histone H2A
1011 variants H2AX and H2AZ. *Curr. Opin. Genet. Dev.* 12, 162–169.

1012 Ritzel, R.M., Patel, A.R., Grenier, J.M., Crapser, J., Verma, R., Jellison, E.R., and McCullough, L.D.
1013 (2015). Functional differences between microglia and monocytes after ischemic stroke. *J.*
1014 *Neuroinflammation* 12, 1–12.

1015 Roddie, H.G., Armitage, E.L., Coates, J.A., Johnston, S.A., and Evans, I.R. (2019). Simu-dependent
1016 clearance of dying cells regulates macrophage function and inflammation resolution. *PLoS Biol.* 17,
1017 e2006741.

1018 Ryglewski, S., Duch, C., and Altenhein, B. (2017). Tyramine Actions on *Drosophila* Flight Behavior Are
1019 Affected by a Glial Dehydrogenase/Reductase. *Front. Syst. Neurosci.* 11, 68.

1020 Santos-Rosa, H., Schneider, R., Bannister, A.J., Sherriff, J., Bernstein, B.E., Emre, N.C.T., Schreiber, S.L.,
1021 Mellor, J., and Kouzarides, T. (2002). Active genes are tri-methylated at K4 of histone H3. *Nature* 419,
1022 407–411.

1023 Schindelin, J., Arganda-Carreras, I., Frise, E., Kaynig, V., Longair, M., Pietzsch, T., Preibisch, S., Rueden,
1024 C., Saalfeld, S., Schmid, B., et al. (2012). Fiji: an open-source platform for biological-image analysis. *Nat.*
1025 *Methods* 9, 676–682.

1026 Schmid, A., Chiba, A., and Doe, C.Q. (1999). Clonal analysis of *Drosophila* embryonic neuroblasts: neural
1027 cell types, axon projections and muscle targets. *Development* 126, 4653–4689.

1028 Shepherd, D. (2000). Glial dependent survival of neurons in *Drosophila*. *Bioessays* 22, 407–409.

1029 Sonnenfeld, M.J., and Jacobs, J.R. (1995a). Macrophages and glia participate in the removal of apoptotic
1030 neurons from the *Drosophila* embryonic nervous system. *J. Comp. Neurol.* 359, 644–652.

1031 Sonnenfeld, M.J., and Jacobs, J.R. (1995b). Macrophages and glia participate in the removal of apoptotic
1032 neurons from the *Drosophila* embryonic nervous system. *J. Comp. Neurol.* 359, 644–652.

1033 Stork, T., Engelen, D., Krudewig, A., Silies, M., Bainton, R.J., and Klambt, C. (2008). Organization and
1034 function of the blood-brain barrier in *Drosophila*. *J. Neurosci.* 28, 587–597.

1035 Syed, A., and Tainer, J.A. (2018). The MRE11-RAD50-NBS1 Complex Conducts the Orchestration of
1036 Damage Signaling and Outcomes to Stress in DNA Replication and Repair. *Annu. Rev. Biochem.* 87, 263–
1037 294.

1038 Tepass, U., Fessler, L.I., Aziz, A., and Hartenstein, V. (1994). Embryonic origin of hemocytes and their
1039 relationship to cell death in *Drosophila*. *Development* 120, 1829–1837.

1040 Thomas, P.D., Campbell, M.J., Kejariwal, A., Mi, H., Karlak, B., Daverman, R., Diemer, K., Muruganujan,
1041 A., and Narechania, A. (2003). PANTHER: A Library of Protein Families and Subfamilies Indexed by
1042 Function. *Genome Res.* 13, 2129–2141.

1043 Trapnell, C., Pachter, L., and Salzberg, S.L. (2009). TopHat: discovering splice junctions with RNA-Seq.
1044 *Bioinformatics* 25, 1105–1111.

1045 Volkenhoff, A., Weiler, A., Letzel, M., Stehling, M., Klämbt, C., and Schirmeier, S. (2015). Glial
1046 Glycolysis Is Essential for Neuronal Survival in *Drosophila*. *Cell Metab.* 22, 437–447.

1047 Wang, Q., Goldstein, M., Alexander, P., Wakeman, T.P., Sun, T., Feng, J., Lou, Z., Kastan, M.B., and
1048 Wang, X.-F. (2014). Rad17 recruits the MRE11-RAD50-NBS1 complex to regulate the cellular response
1049 to DNA double-strand breaks. *EMBO J.* 33, 862–877.

1050 Watts, R.J., Schuldiner, O., Perrino, J., Larsen, C., and Luo, L. (2004). Glia engulf degenerating axons
1051 during developmental axon pruning. *Curr. Biol.* *14*, 678–684.

1052 Wu, Y., Dissing-Olesen, L., MacVicar, B.A., and Stevens, B. (2015). Microglia: Dynamic Mediators of
1053 Synapse Development and Plasticity. *Trends Immunol.* *36*, 605–613.

1054 Zuber, R., Wang, Y., Gehring, N., Bartoszewski, S., and Moussian, B. (2020). Tweedle proteins form
1055 extracellular two-dimensional structures defining body and cell shape in *Drosophila melanogaster*. *Open*
1056 *Biol.* *10*, 200214.

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1058

1059 **Figure legends**

1060 **Figure 1: RNAseq analysis in embryonic and larval glia, neurons and hemocytes**

1061 (a) Dendrogram showing the hierarchical clustering based on the distance between samples. The
1062 height indicates the distance. Euclidean distance, Ward criterion were used for this representation.

1063 (b-c) Venn diagrams showing the distribution of differentially expressed genes between neurons
1064 (blue), glia (yellow) and hemocytes (red) at E16 (b) and L3 (c). Since only upregulated genes are
1065 used for the analysis, the intersection between all three cell types is empty (white). The percentage
1066 of genes in each category is indicated in the diagram. The percentage was calculated based on all
1067 genes used for the analysis. (d,e) The total number of genes expressed (top three lines) and
1068 upregulated compared the other two cell types (bottom three lines) at E16 (d) and L3 (e).

1069 **Figure 2: Pairwise transcriptome comparisons in the embryo**

1070 (a) Transcriptome comparison of E16 neurons and glia, MA plot. The x-axis reports the average
1071 gene expression levels, the y-axis, the log₂ fold change between the genes in E16 neurons/E16
1072 glia. Genes significantly upregulated in E16 neurons are shown in blue, genes significantly
1073 upregulated in E16 glia are shown in yellow and non-differentially expressed genes are shown in
1074 gray. (b,c) Gene Ontology (GO) term enrichment analysis in E16 neurons (blue) and E16 glia
1075 (yellow). The fold enrichment for a subset of representative GO terms is displayed on the x-axis,
1076 the number of genes within the GO term and the P-value of the GO term enrichment are indicated
1077 in brackets. The complete results of the GO term analysis are shown in **Sup. Table 2-b,c**. (d)
1078 Transcriptome comparison of E16 hemocytes and glia. MA plot as in (a), the y-axis reports the
1079 log₂ fold change E16 hemocytes/E16 glia. Genes significantly upregulated in E16 hemocytes are
1080 shown in red, genes significantly upregulated in E16 glia are shown in yellow and non-
1081 differentially expressed genes are shown in gray. (e,f) GO term enrichment analysis in E16

1082 hemocytes (red) and E16 glia (yellow). The fold enrichment for a subset of representative GO
1083 terms is displayed on the x-axis, the number of genes and the P-value of the GO term enrichment
1084 are indicated in brackets. The complete results of the GO term analysis are shown in **Sup. Table**
1085 **2-d,e.** (g) List of genes upregulated in E16 hemocytes compared to E16 glia and involved in the
1086 immune deficiency and Toll pathways. (h) Transcriptome comparison of E16 neurons and
1087 hemocytes. MA plot as in (a), the y-axis reports the log₂ fold change E16 neurons/E16 hemocytes.
1088 Genes significantly upregulated in E16 neurons are shown in blue, genes significantly upregulated
1089 in E16 hemocytes in red, and non-differentially expressed genes in gray.

1090 **Figure 3: Pairwise transcriptome comparisons in the larva**

1091 (a) Transcriptome comparison of L3 neurons and glia, MA plot. The x-axis reports the average
1092 gene expression levels, the y-axis, the log₂ fold change L3 neurons/ L3 glia. Genes significantly
1093 upregulated in L3 neurons are shown in blue, genes significantly upregulated in L3 glia are shown
1094 in yellow and non-differentially expressed genes are shown in gray. (b,c) Gene Ontology (GO)
1095 term enrichment analysis in L3 neurons (blue) and L3 glia (yellow). The fold enrichment for a
1096 subset of representative GO terms are displayed on the x-axis, the number of genes and the P-value
1097 of the GO term enrichment are indicated in brackets. The complete results of the GO term analysis
1098 are shown in **Sup. Table 3-a,b.** (d) Transcriptome comparison of L3 hemocytes and glia. MA plot
1099 as in (a), the y-axis reports the log₂ fold change L3 hemocytes/L3 glia. Genes significantly
1100 upregulated in L3 hemocytes are shown in red, genes significantly upregulated in L3 glia are
1101 shown in yellow and non-differentially expressed genes are shown in gray. (e,f) Gene Ontology
1102 (GO) term enrichment analysis in L3 hemocytes (red) and L3 glia (yellow). The fold enrichment
1103 for a subset of representative GO terms is displayed on the x-axis, the number of genes and the P-
1104 value of the GO term enrichment are indicated in brackets. The complete results of the GO term
1105 analysis are shown in **Sup. Table 3-c,d.** (g) Transcriptome comparison of L3 neurons and
1106 hemocytes. MA plot as in (a), the y-axis reports the log₂ fold change L3 neurons/L3 hemocytes.
1107 Genes significantly upregulated in L3 neurons are shown in blue, genes significantly upregulated
1108 in L3 hemocytes are shown in red, and non-differentially expressed genes are shown in gray.

1109 **Figure 4: *NimA*, a novel glia-specific scavenger receptor**

1110 (a,b) MA plots as in Figure 2-c and Figure 3-c comparing the scavenger receptors expressed in
1111 embryonic and larval hemocytes and glia. In black, scavenger receptors non-differentially

1112 expressed between E16 (a) or L3 hemocytes and glia (b). In red, scavenger receptors that are
1113 upregulated in E16 (a) or L3 hemocytes (b) and in yellow, scavenger receptors that are upregulated
1114 in E16 (a) or L3 glia (b). (c,e') Immunolabeling of *NimAGal4;UAS-eGFP* embryos (c,c'), L3
1115 (d,d',d'') and adults (e) with anti GFP (green), anti-Repo (white), anti-Nazgul (red) and DAPI
1116 (blue). (c) Full stack projection, scale bar = 100 μm . Square indicates region in c'. (c') Single
1117 section, scale bar = 50 μm . (d) Full stack projection, scale bar = 100 μm . Squares indicate regions
1118 in d' and d''. (d'-d'') Single section, scale bar = 50 μm . (e) Full stack projection, scale bar = 100
1119 μm .

1120 **Figure 5: Pairwise transcriptome comparison between E16 and L3 cells**

1121 (a) Transcriptome comparison of L3 vs. E16 cells, MA plot. The x-axis reports the average gene
1122 expression levels, the y-axis, the log₂ fold change L3 cells/E16 cells. Genes significantly
1123 upregulated in E16 are shown in green, genes significantly upregulated in L3 are shown in purple
1124 and non-differentially expressed genes are shown in gray. (b,d) GO term enrichment analysis in
1125 L3 (purple) and E16 (green). The fold enrichment for a subset of representative GO terms is
1126 displayed on the x-axis, the number of genes and the P-value of the GO term enrichment are
1127 indicated in brackets. The complete results of the GO term analysis are shown in **Sup. Table 6-**
1128 **a,c**. (c,e,f) Heatmaps showing the log₁₀ gene expression of genes upregulated in E16 cells
1129 compared to L3 cells and involved in Chitin-based cuticle development (c), as well as that of genes
1130 upregulated in L3 cells compared to E16 cells and involved in DNA repair (e) and epigenetics (f).
1131 Scale shows low expression (blue) to high expression (red). Values for heatmaps shown in **Sup.**
1132 **Table 6-b,d,e**.

1133 **Figure 6: Impact of *rad50* on polytene chromosomes and on gene expression**

1134 (a) Representative polytene chromosomes immunolabeled with anti-H3K4me₃ (magenta) and
1135 anti-H3K9me₂ (green), or with anti-H3K9me₃ (red), from *WT* or *rad50 Δ 5.1* homozygous mutants
1136 L3. Chromosomes were counter-stained with DAPI (blue). Scale bar = 20 μm . (b) Fluorescence
1137 intensity quantification of polytene chromosomes, labeled as in (a), shows an increase of the
1138 euchromatic marker H3K4me₃ and a decrease of the heterochromatic markers H3K9me₂
1139 H3K9m₃, in *rad50 Δ 5.1* mutants compared to *WT*. Each dot represents a single polytene
1140 chromosome (n \geq 13). Arbitrary Unit (A.U.). Error bars represent SEM. P = p-value calculated
1141 using Welch parametric t-test. (c) Representative intensity profiles of H3K4me₃ and DAPI signals

1142 over a segment at the tip of chromosome 2L, showing an alteration of the H3K4me3 signal in
1143 *rad50Δ5.1* mutant compared to the *WT*; arrowheads denote regions of strong difference of
1144 H3K4me3 in *rad50Δ5.1* compared to *WT*. y-axis: fluorescence intensity; x-axis: distance from the
1145 tip of the chromosome (μm). Scale bar = 10 μm. (d) Representative intensity profiles of H3K9me3
1146 and DAPI signals over a segment at the tip of chromosomes 3R showing absence of H3K9me3
1147 signal in *rad50Δ5.1* compared to *WT*. y-axis: fluorescence intensity; x-axis: distance from the tip
1148 of the chromosome (μm). Scale bar = 10 μm. (e) Representative polytene chromosomes
1149 immunolabeled with anti-H3K27me3 (purple) and anti-H2A.v-P (green), from *WT* or *rad50Δ5.1*
1150 homozygous larvae. Chromosomes were counter-stained with DAPI (blue). Scale bar = 20 μm. (f)
1151 Fluorescence intensity quantification showing increase of both H3k27me3 and H2A.v-P signals in
1152 *rad50Δ5.1* mutants compared to *WT*. Each dot represents a single polytene chromosome (n ≥
1153 15). Error bars represent SEM. P = p-value calculated using Welch parametric t-test. (g)
1154 Representative intensity profiles of H3K27me3 and H2A.v-P signals over a segment at the tip of
1155 the 2R chromosome. Note that the increase of H2A.v-P peaks in the *rad50Δ5.1* mutant is not
1156 directly correlated with that of H3K27me3 (arrowhead). y-axis: fluorescence intensity; x-axis:
1157 distance from the tip of the chromosome (μm). Scale bar = 10 μm. (e-j) RT-qPCR assays on
1158 *rad50Δ5.1* /+ and *rad50Δ5.1* larvae. The levels of *TwdlW* (h), *ftz-fl* (i), *mre11* (j), *Unc-76* (k),
1159 *olf413* (l) and *sna* (m) transcripts were quantified and the relative expression levels were calculated
1160 based on the expression of *Act5C* and *RP49*. List of primers shown in **Sup. Table 9**, n = number
1161 of biological replicates, P = p-value calculated using Student's t-test and indicated on graphs, error
1162 bars represent the standard deviation.

1163 **Figure 7: Comparison of the metabolic states in hemocytes, glia and neurons at E16 and L3**
1164 (a) Schematic showing the main steps of sugar and lipid metabolic pathways. (b,c) Heatmaps
1165 showing the z-score of genes involved in the different metabolic processes (b) and transporters (c)
1166 in hemocytes, glia and neurons at E16 and L3. Scale shows low expression (blue) to high
1167 expression (red). Values are shown in **Sup. Table 7**. (d) Recapitulative table highlighting the main
1168 processes for sugar and lipid metabolism in hemocytes, neurons and glia from E16 and L3.

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1170

1171 **Supplementary figure legends**

1172 **Sup. Figure 1: Read counts and commonalities between embryonic neurons and glia**

1173 (a) Comparison of the total read count between the different samples. The y-axis represents the
1174 total read count. The x axis-represents the different samples: E16 glia, neurons and hemocytes
1175 (blue) and L3 glia, neurons and hemocytes (red). (b,c) Dot plots comparing the expression levels
1176 of non-differentially genes versus differentially expressed genes. y-axis reports the log₂ fold
1177 change E16 neurons/E16 glia. Non-differentially expressed genes are shown in gray (b). The x-
1178 axis reports the average gene expression levels. Genes significantly upregulated in E16 neurons
1179 are shown in blue, genes significantly upregulated in E16 glia are shown in yellow (c). (d) Gene
1180 Ontology (GO) term enrichment analysis on genes commonly expressed between E16 glia and
1181 neurons. The fold enrichment for a subset of representative GO terms is displayed on the x-axis,
1182 the number of genes and the P-value of the GO term enrichment are indicated in brackets. The
1183 complete results of the GO term analysis are shown in **Sup. Table 2-a**.

1184 **Sup. Figure 2: Cell type differences between E16 and L3**

1185 (a) Box plot showing the log₂ fold change (y-axis) between the different samples at E16 and L3
1186 (x-axis). P = p-value calculated using Student's t-test and indicated on graphs, error bars represent
1187 the standard deviation. (b) Heatmap showing the Log₁₀ expression levels (absolute levels) of
1188 genes upregulated in L3 hemocytes and L3 glia but not in neurons and involved in metabolism.
1189 Scale shows low expression (blue) to high expression (red). Log₁₀ expression values are shown
1190 in **Sup. Table 4-b,c**. (c-e) Transcriptome comparison of the cell types at E16 vs. L3. The x-axis
1191 reports the average gene expression levels, the y-axis, the log₂ fold change of L3 glia / E16 glia
1192 (c), L3 neurons / E16 neurons (d) and L3 hemocytes / E16 hemocytes (e). Genes significantly
1193 upregulated in L3 glia, L3 neurons and L3 hemocytes are shown in orange, dark blue and dark red
1194 respectively. Genes significantly upregulated in E16 glia, E16 neurons and E16 hemocytes are
1195 shown in yellow, light blue and light red, respectively. Non-differentially expressed genes are
1196 shown in gray. (f,g) Venn diagrams showing the distribution of transcription factors between E16
1197 neurons (blue), glia (yellow) and hemocytes (red) (f), and L3 neurons (blue), glia (yellow) and
1198 hemocytes (red) (g). Transcription factors common to all three cell types are shown in white. The
1199 number of transcription factors in each category is indicated in the diagram.

1200 **Sup. Figure 3: Commonalities and specificities between hemocytes and glia**

1201 (a) Graphical representation of NimA and Drpr. (b-h) Quantitative reverse transcription PCR (RT-
1202 qPCR) assays on sorted L3 glia and hemocytes. To test the purity of the sorted populations, the
1203 expression of *Hml* (b) and *repo* (c) were assessed in hemocytes and glia, respectively. The levels
1204 of *NimC1* (d), *eater* (e), *drpr* (f), *crq* (g) and *NimA* (h) were quantified and relative expression
1205 levels were calculated based on the expression of *Act5C* and *RP49*. List of primers shown in **Sup.**
1206 **Table 9**, n = number of biological replicates, P = p-value calculated using Student's t-test and
1207 indicated on graphs, error bars represent the standard deviation. NS = non-significant. (i) *In situ*
1208 hybridization using an anti-*NimA* RNAscope probe on L3 CNS, full stack projection. Region in
1209 white square in (i) is shown magnified in (i'). *NimA* is shown in red, DAPI in blue. (i) Scale bar =
1210 100 μ m, (i') Scale bar = 50 μ m.

1211 **Sup. Figure 4: Specificities of larval cells**

1212 (a-c) RT-qPCR assays on cells from E16 and L3 *WT* animals. The levels of *rad50* (a), *mre11* (b)
1213 and *nbs* (c) transcripts were quantified and the relative expression levels were calculated based on
1214 the expression of *Act5C* and *RP49*. List of primers shown in **Sup. Table 9**, n = number of
1215 biological replicates, p-value calculated using Student's t-test and indicated on graphs, error bars
1216 represent the standard deviation. (d,e) Western blot quantification of γ H2AV. Histones extracted
1217 from cells from E16 and L3 *WT* animals were blotted on nitrocellulose membranes and probed
1218 with anti- H2A.v-P antibody (e). An antibody that recognizes all histones H3, anti-H3, was used
1219 to confirm equal loading of lysates (d). The ladder on the left shows protein size in KDa. (f) Cells
1220 were first selected based on the forward scatter (FSC-A) and the side scatter (SSC-A) to eliminate
1221 dead cells and debris. Doublets were removed and only single cells were taken into account. The
1222 gate for sorting Cy3/RFP positive cells was based on the negative control. Only Cy3/RFP positive
1223 were sorted. A post sort step was performed at the end to test the purity of the sorted population.

1224 **Sup. Figure 5: Comparison of the metabolic pathways expressed in the three cell types at**
1225 **embryonic and larval stages**

1226 (a,b) Heatmaps showing the Log₁₀ expression levels (absolute levels) of genes involved in the
1227 main metabolic processes (a) and of the main transporters (b) in hemocytes, glia and neurons of
1228 E16 and L3. Scale shows low expression (blue) to high expression (red). Expression values are
1229 shown in **Sup. Table 7**.

1230 **List of supplementary tables:**

1231 **Sup. Table 1: List of transcription factors and their expression levels in E16 and L3 glia,**
1232 **neurons and hemocytes.**

1233 **Sup. Table 2: Gene Ontology terms found in commonly expressed genes or genes upregulated**
1234 **in E16 glia, E16 neurons or E16 hemocytes.**

1235 **Sup. Table 3: Gene Ontology terms found in genes upregulated in L3 glia, L3 neurons or L3**
1236 **hemocytes.**

1237 **Sup. Table 4: Gene Ontology terms and list of genes upregulated in L3 glia and hemocytes**
1238 **compared to neurons.**

1239 **Sup. Table 5: List of scavenger receptors expressed in glia and hemocytes.**

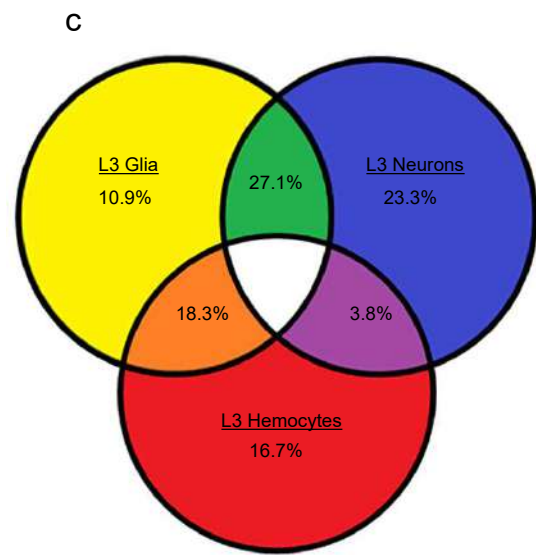
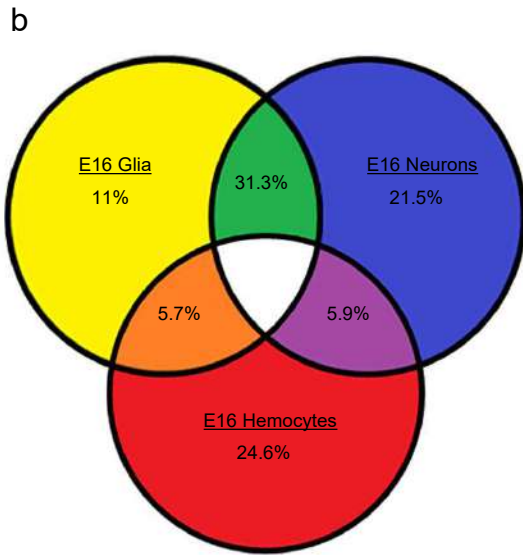
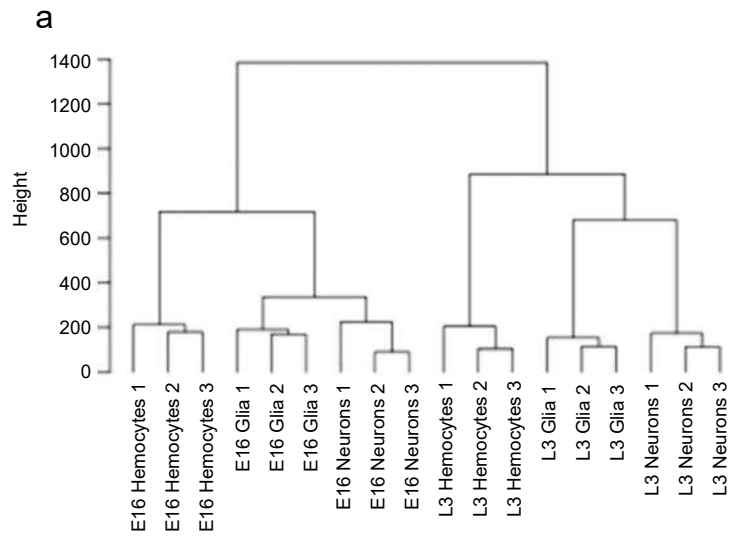
1240 **Sup. Table 6: Gene Ontology terms and lists of genes upregulated in E16 cells or L3 cells.**

1241 **Sup. Table 7: Lists of genes involved in different metabolism pathways.**

1242 **Sup. Table 8: Gene Ontology terms found in genes upregulated in E16 hemocytes or E16**
1243 **neurons.**

1244 **Sup. Table 9: List of primers used in this study.**

Figure 1



d

E16

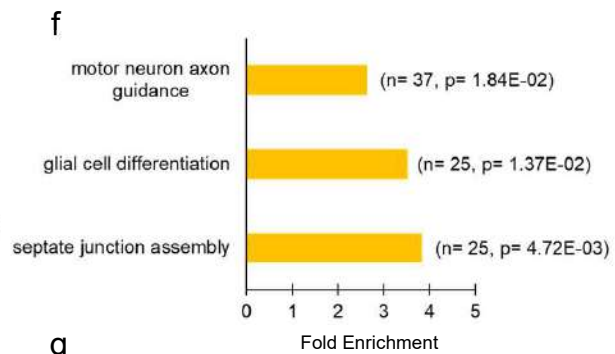
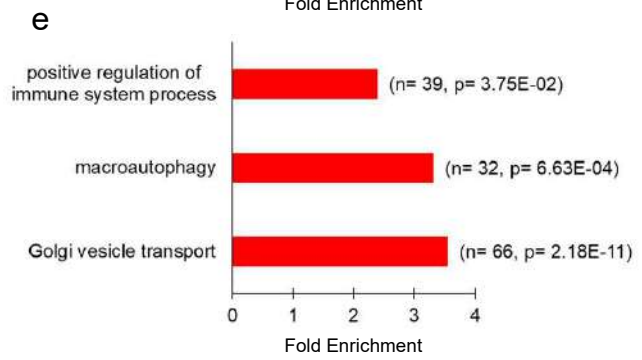
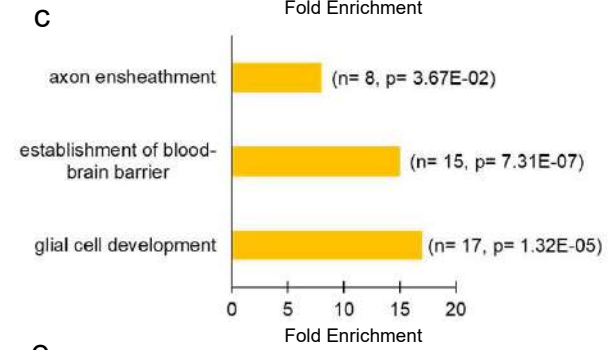
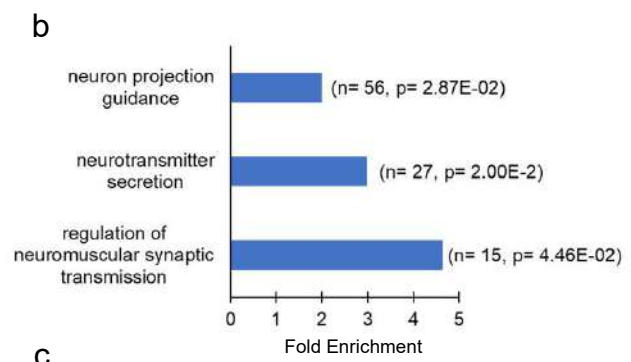
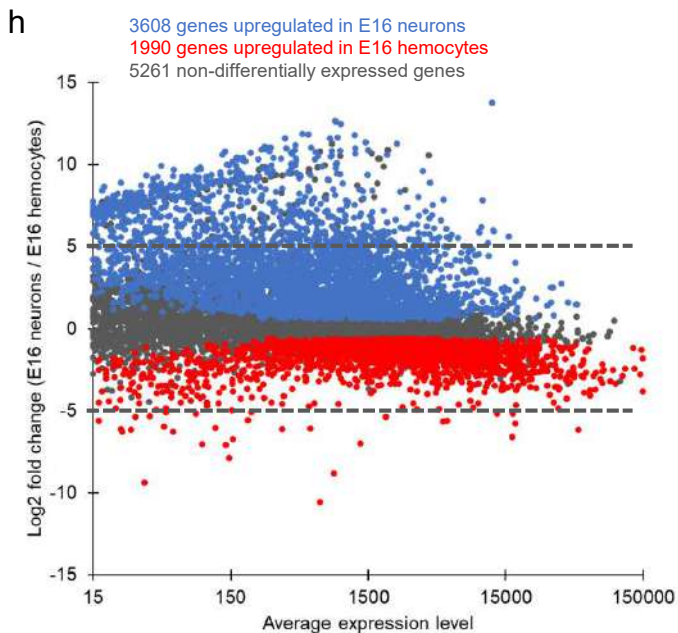
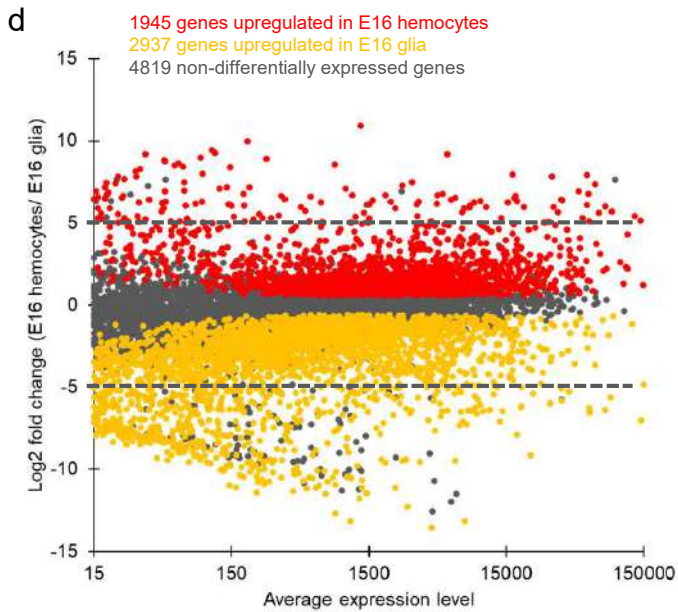
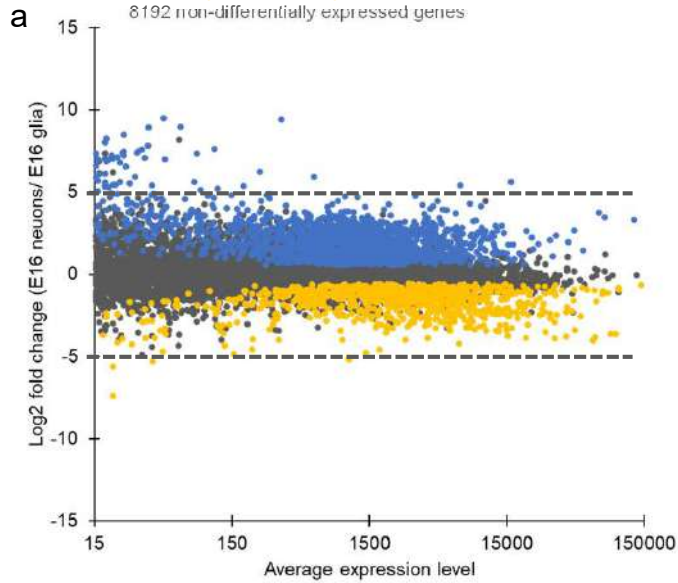
# of genes expressed in E16 glia	10838
# of genes expressed in E16 neurons	11281
# of genes expressed in E16 hemocytes	9811
# of genes upregulated in E16 glia	3414
# of genes upregulated in E16 neurons	4177
# of genes upregulated in E16 hemocytes	2569

e

L3

# of genes expressed in L3 glia	9187
# of genes expressed in L3 neurons	9312
# of genes expressed in L3 hemocytes	7828
# of genes upregulated in L3 glia	4352
# of genes upregulated in L3 neurons	4191
# of genes upregulated in L3 hemocytes	2996

Figure 2 1649 genes upregulated in E16 neurons
992 upregulated in E16 glia
8192 non-differentially expressed genes



g

Immune Deficiency pathway		
FBgn0024222	I-kappaB kinase beta	IKKbeta
FBgn0013983	immune deficiency	imd
FBgn0041205	kenny	key
FBgn0035976	Peptidoglycan recognition protein LC	PGRP-LC
FBgn0031317	Charon	Charon
FBgn0010247	Poly-(ADP-ribose) polymerase	Parp

Toll pathway		
FBgn0011274	Dorsal-related immunity factor	Dif
FBgn0040323	Gram-negative bacteria binding protein 1	GNBP1
FBgn0030310	Peptidoglycan recognition protein SA	PGRP-SA
FBgn0010441	pelle	pll
FBgn0039102	Spatzle-Processing Enzyme	SPE
FBgn0003495	spatzle	spz
FBgn0262473	Toll	Tl
FBgn0034246	Dicer-2	Dcr-2
FBgn0265998	Darkener of apricot	Doa
FBgn0035207	HECT and RLD domain containing E3 ubiquitin ligase 4	Herc4
FBgn0010602	lesswright	lwr
FBgn0051217	modular serine protease	modSP
FBgn0030926	persephone	psh
FBgn0031676	senju	senju

Figure 3 2500 genes upregulated in L3 neurons
2279 genes upregulated in L3 glia
4756 genes non-differentially expressed

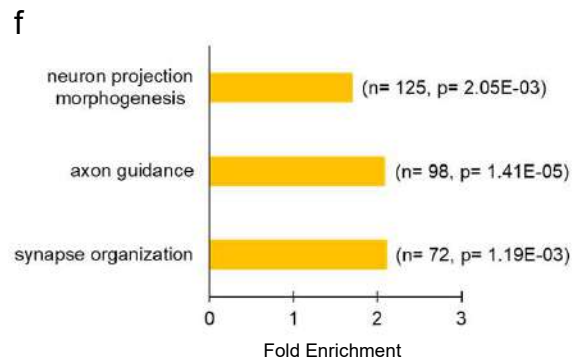
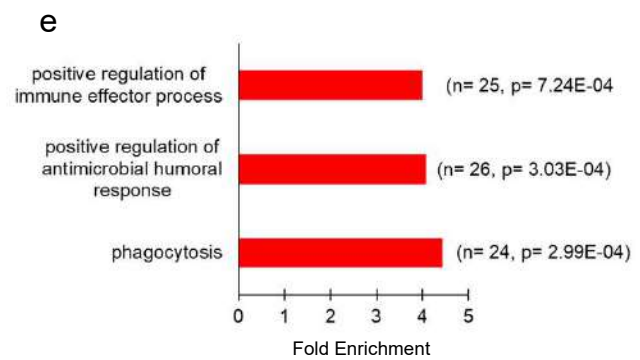
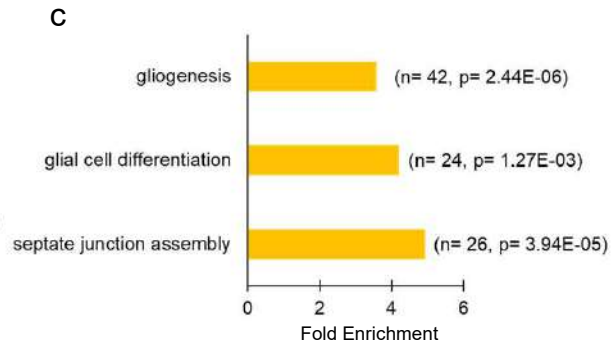
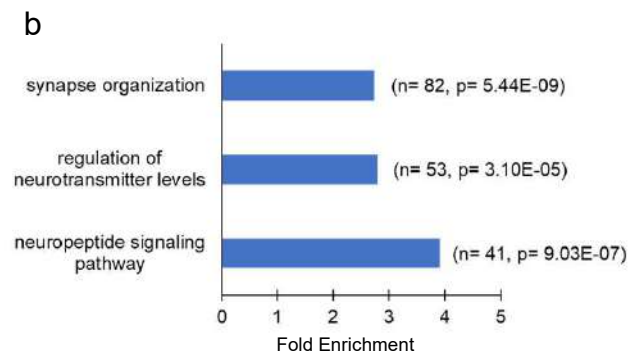
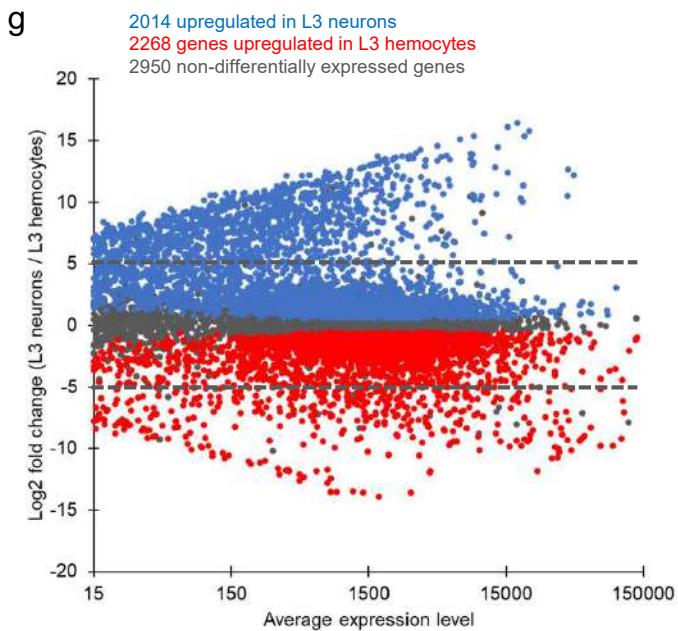
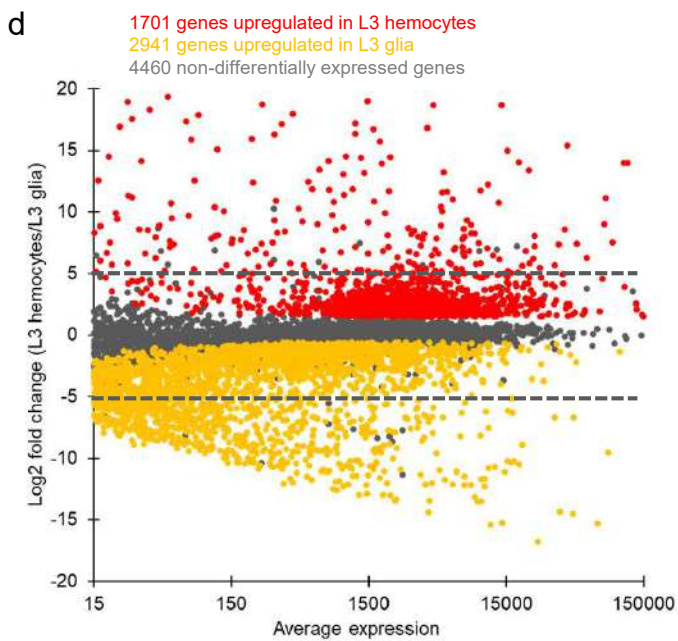
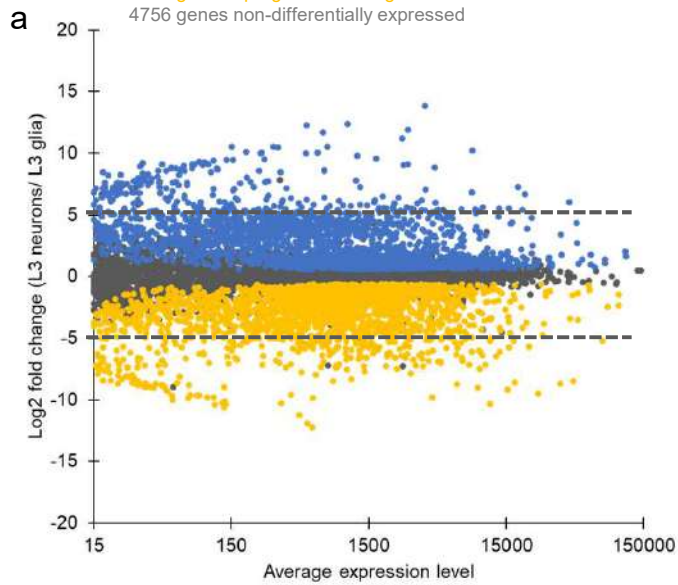
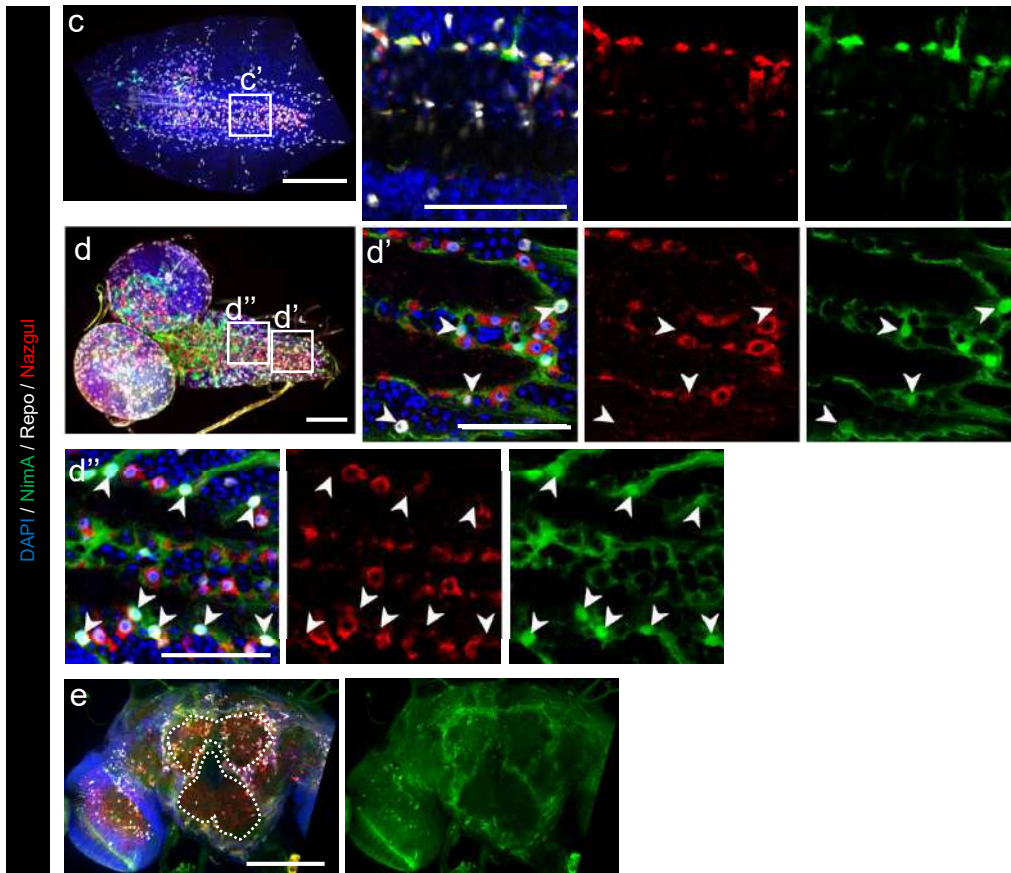
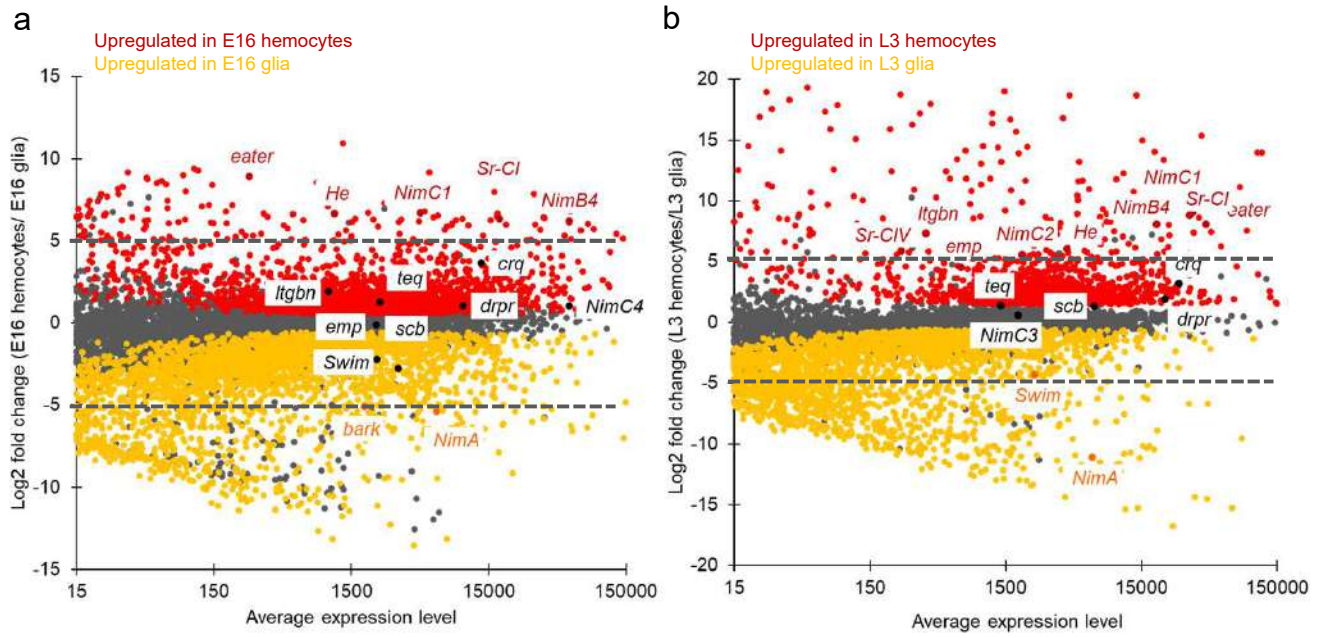


Figure 4



Adult brain scRNAseq
(NimA / ensheathing glia / astrocyte-like glia)

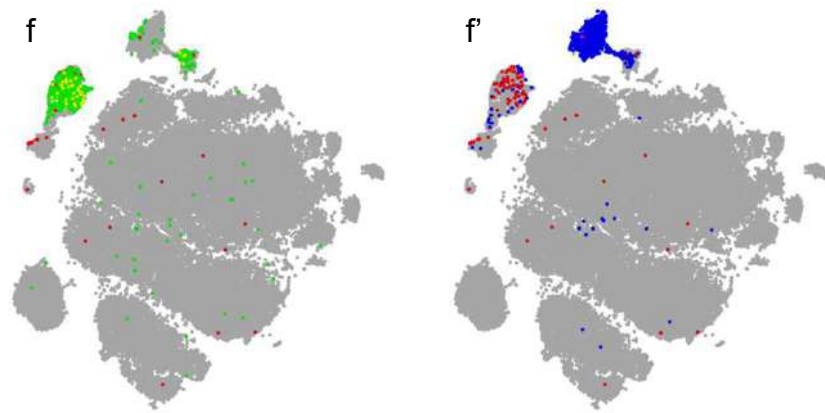


Figure 5

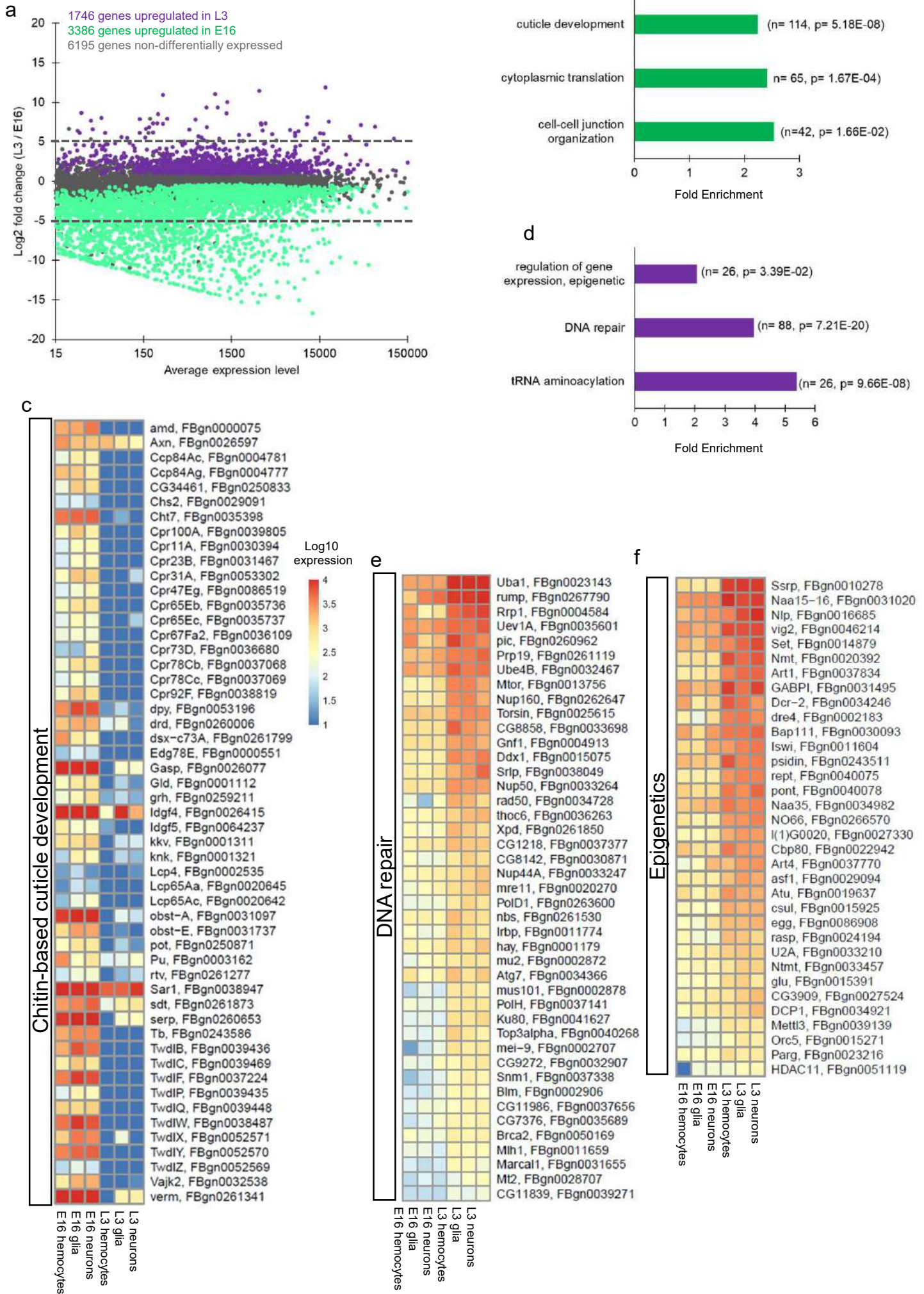


Figure 6

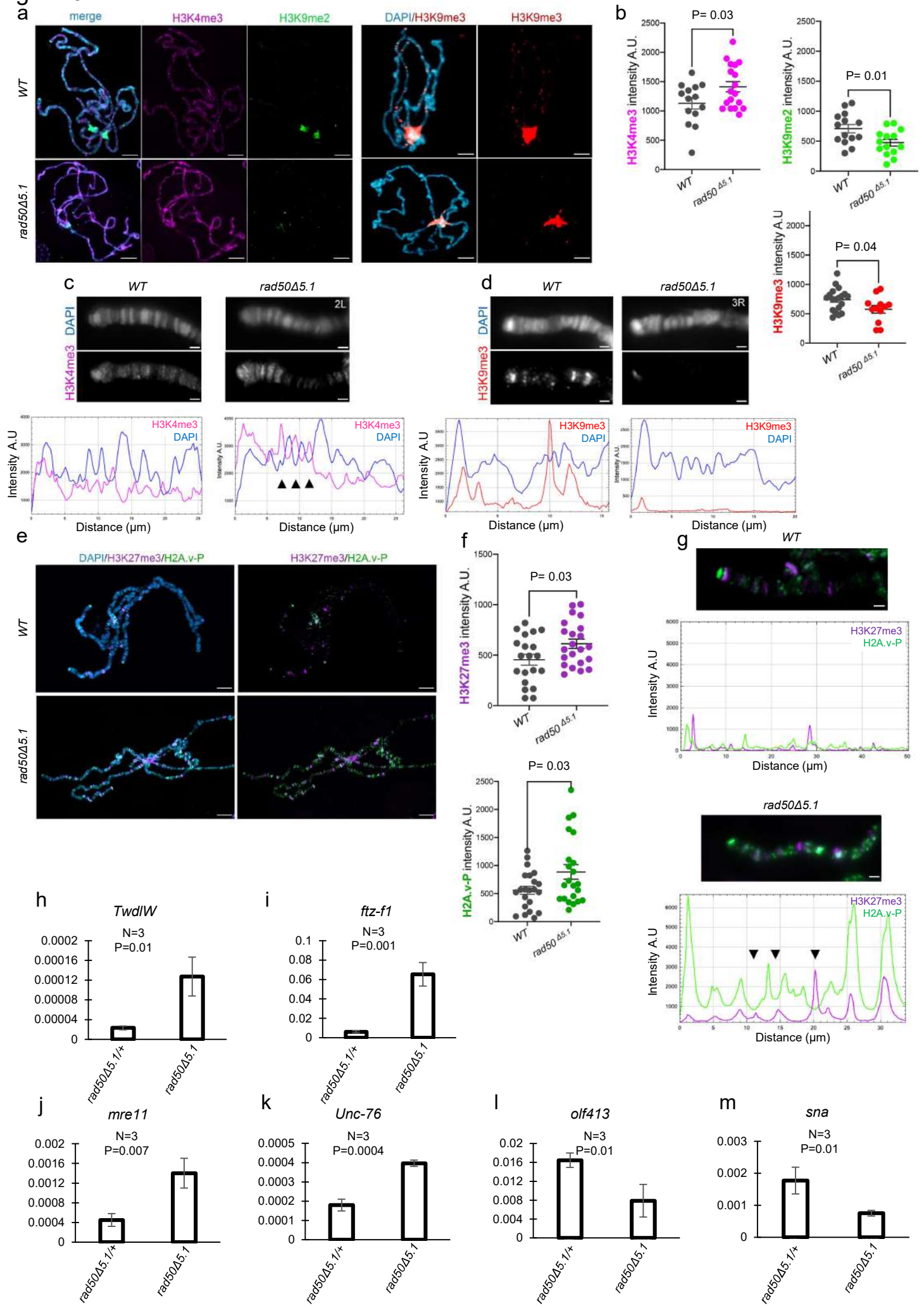
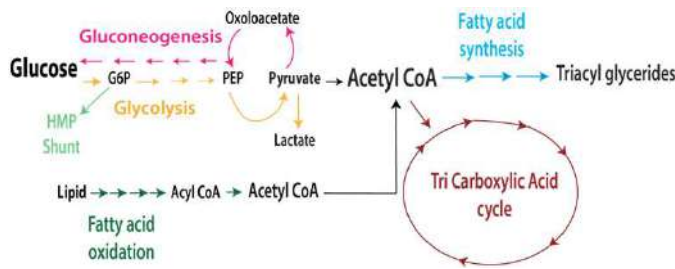
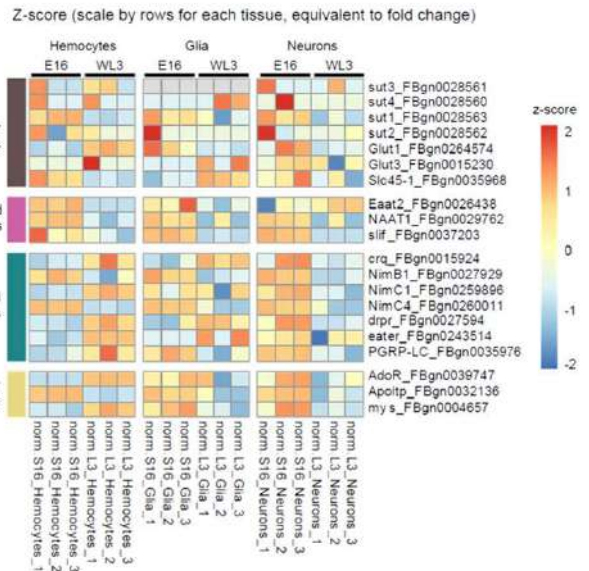


Figure 7

a

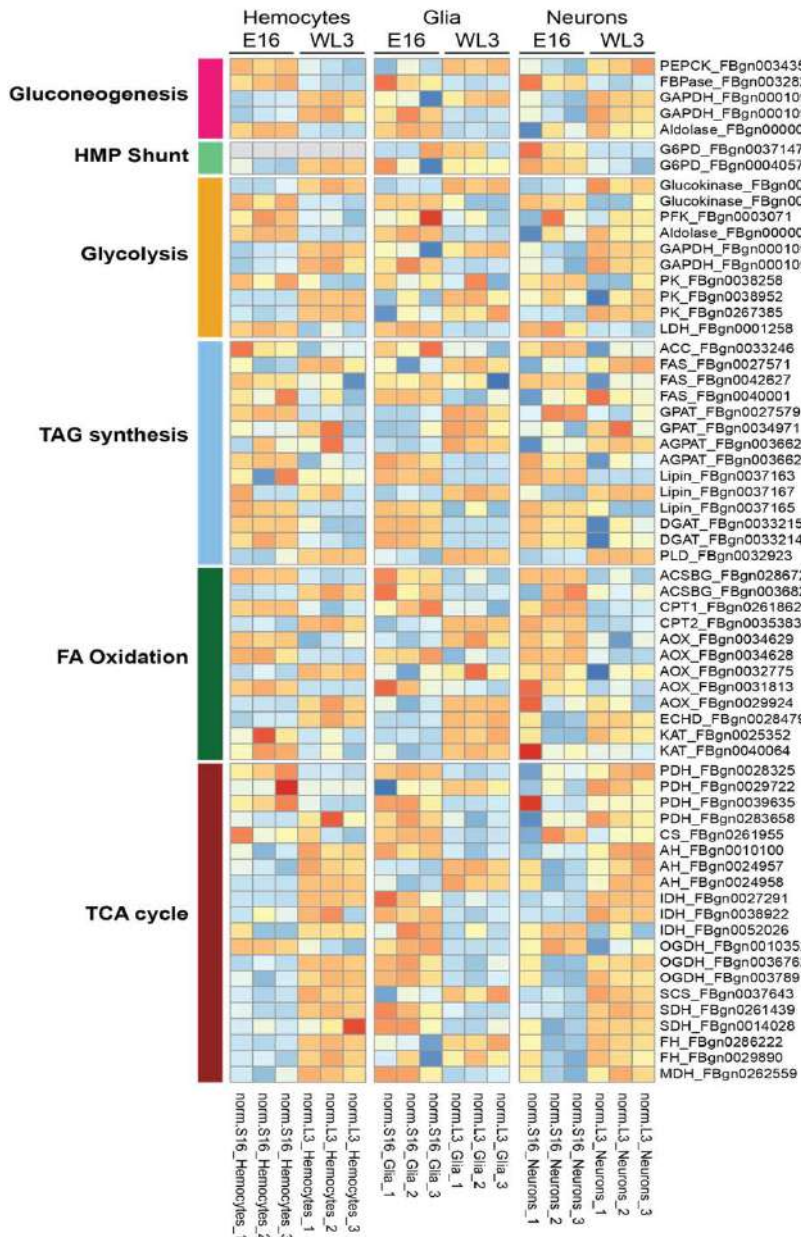


c



b

Z-score (scale by rows for each tissue, equivalent to fold change)



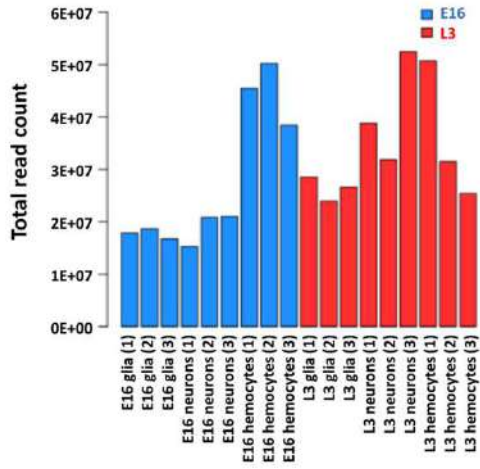
d

Sugar metabolism			
	Hemocytes	Neurons	Glia
Embryo	Gluconeogenic	Glycolytic	Glycolytic
	Glycolytic		Oxidative
Larva	Oxidative	Gluconeogenic	Gluconeogenic
		Oxidative	Oxidative

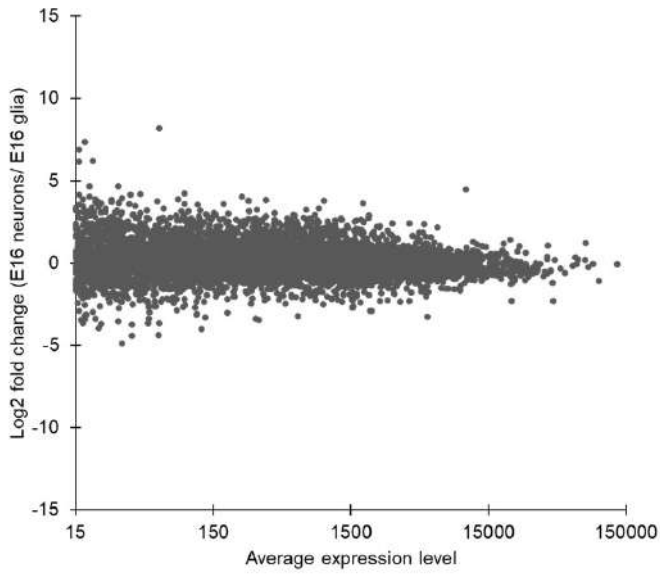
Lipid metabolism			
	Hemocytes	Neurons	Glia
Embryo	Lipogenic	Lipogenic	Lipogenic
		Lipolytic	
Larva	Lipolytic	Lipogenic	Lipogenic
		Lipolytic	Lipolytic

Sup. Figure 1:

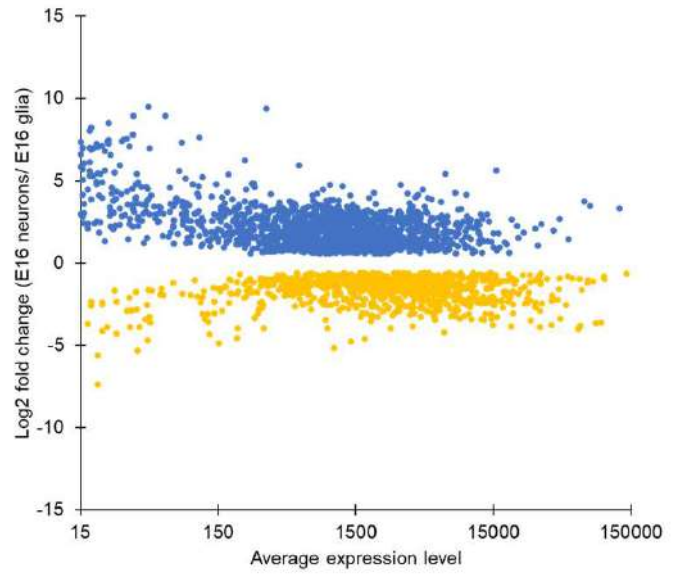
a



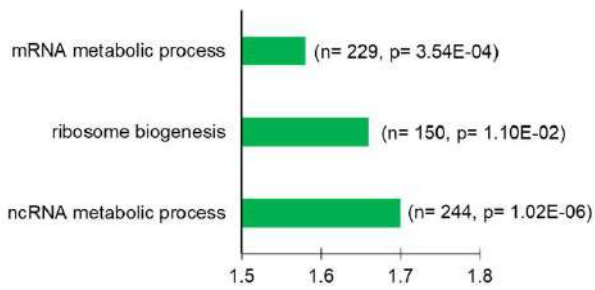
b



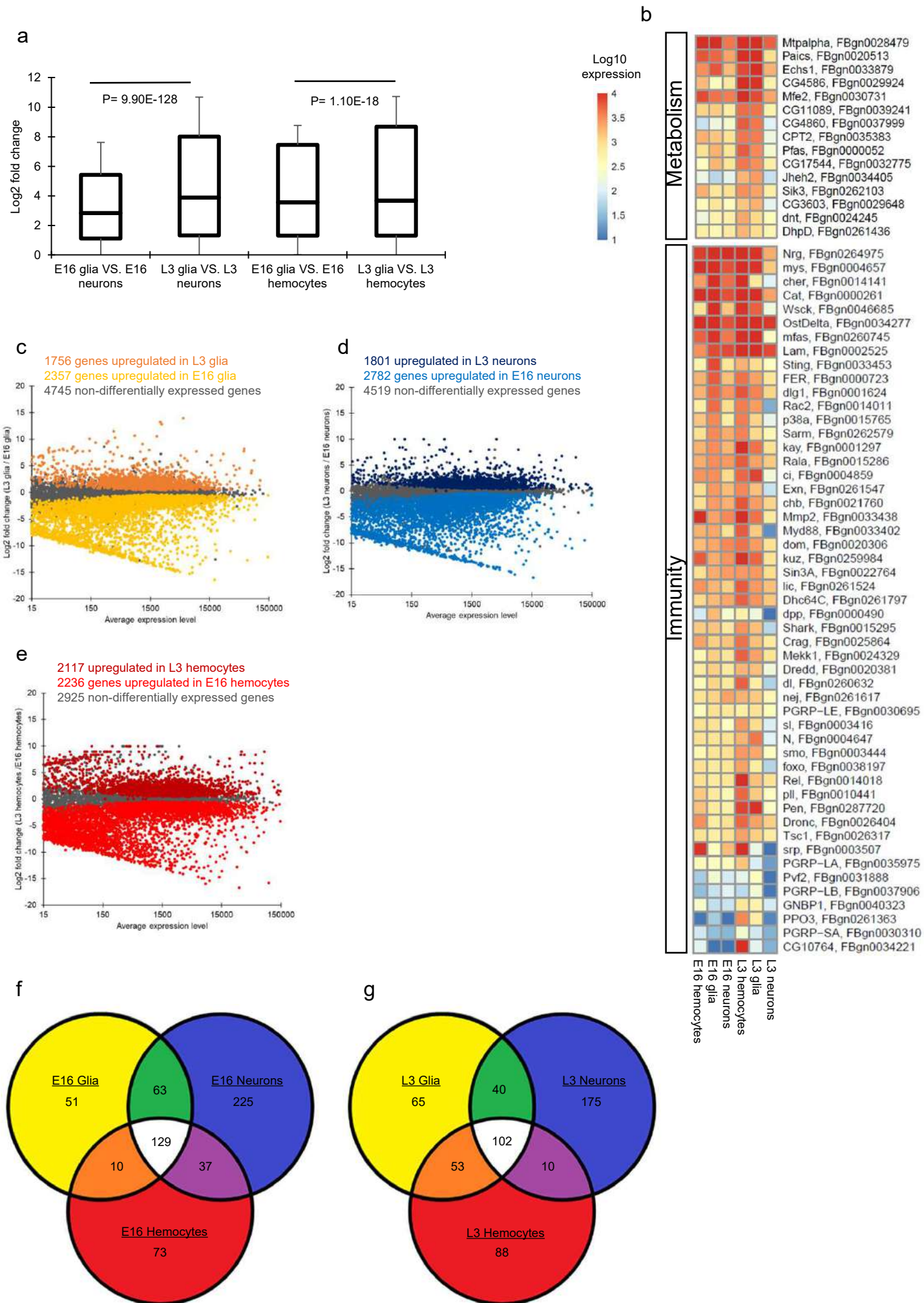
c



d



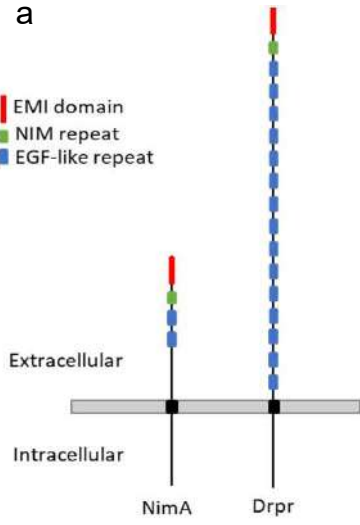
Sup. Figure 2:



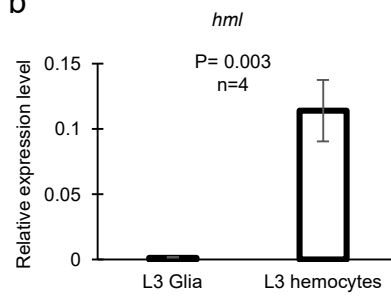
Sup. Figure 3:

a

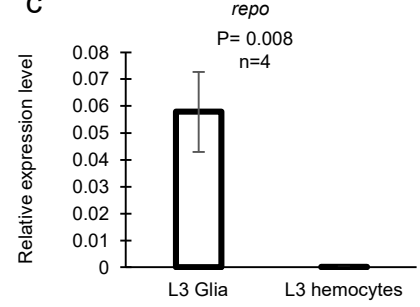
EMI domain
NIM repeat
EGF-like repeat



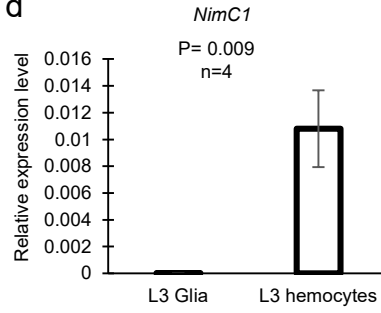
b



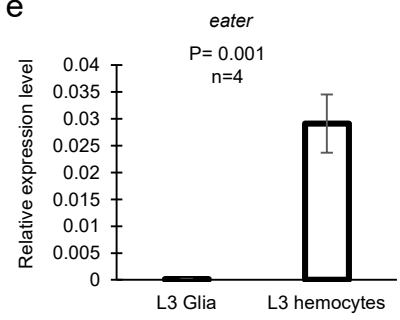
c



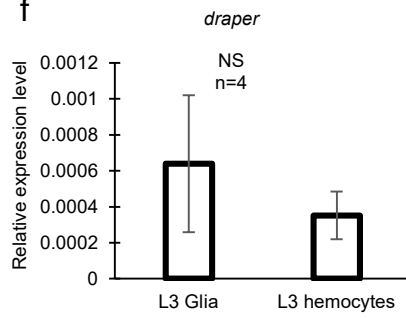
d



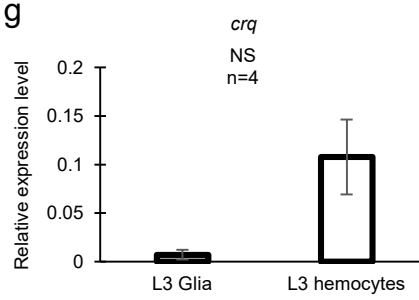
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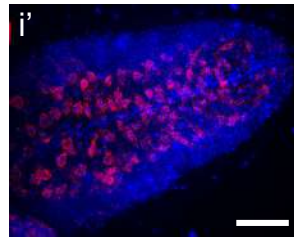
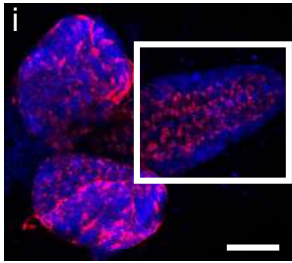
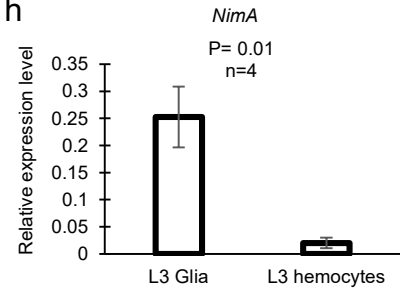
f



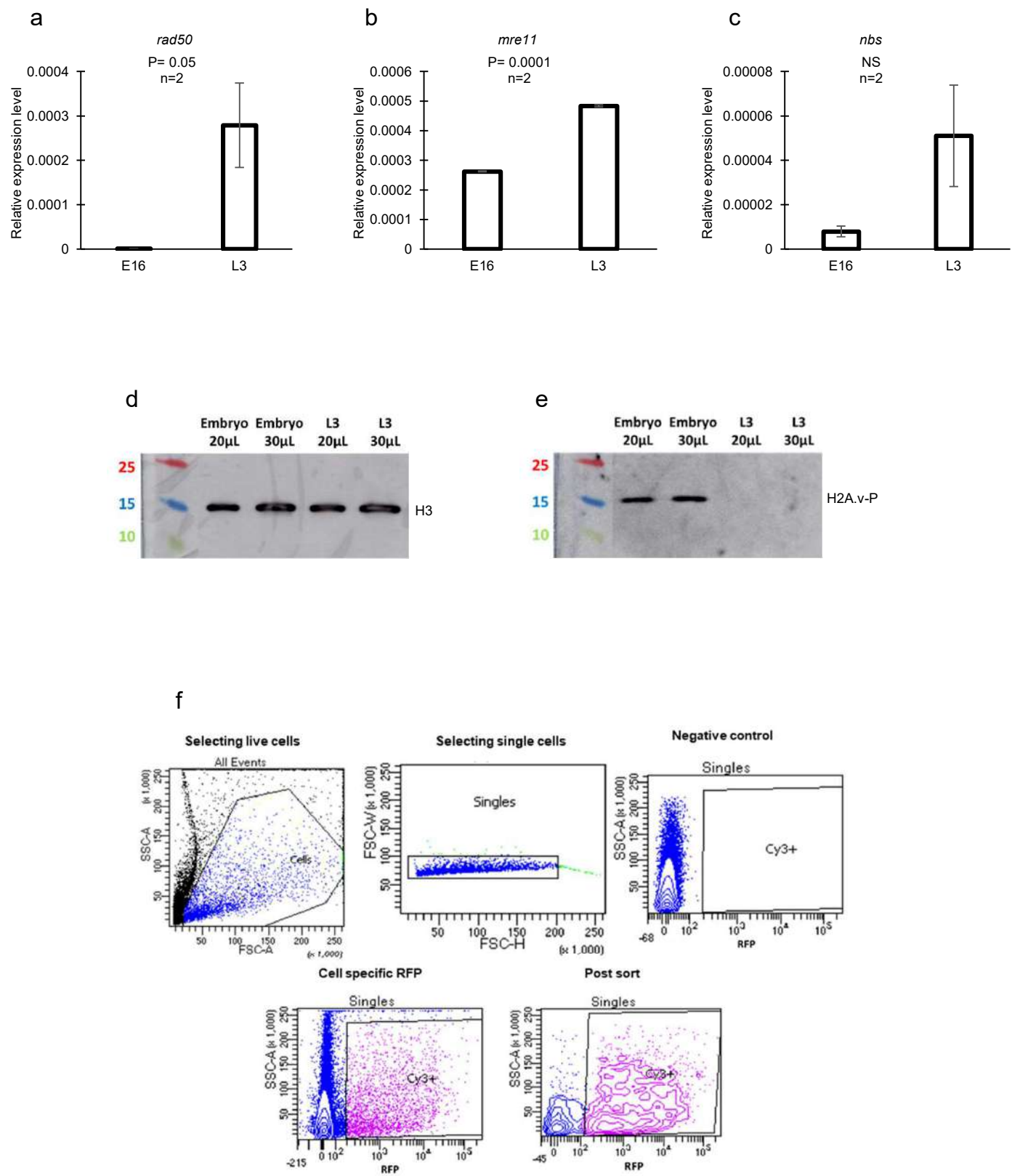
g



h



Sup. Figure 4:



Sup. Figure 5:

